

Machine Learning for Data Generated from Next-Generation Sequencing

DOE-NCI partnership to advance exascale development through cancer research

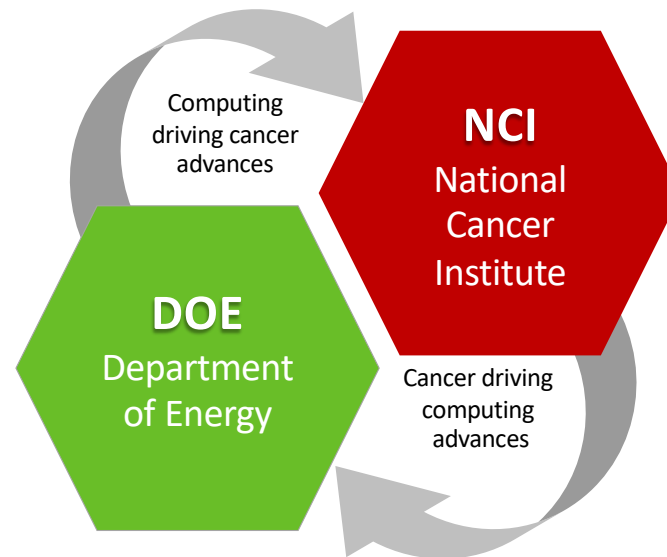
Fangfang Xia

fangfang@anl.gov

Computer Scientist

Argonne National Laboratory

2019-10-23





Jeremy Howard @jeremyphoward · 22h

Seems like there is a real revolution going on in protein analysis thanks to deep learning language models. Here's an example that just got published



Unified rational protein engineering with sequence-...

UniRep learns fundamental protein features from millions of amino-acid sequences using a recurrent neural network. This summary of features can then be used ...

[nature.com](https://www.nature.com)

8

146

629

Representation learning

dnaK

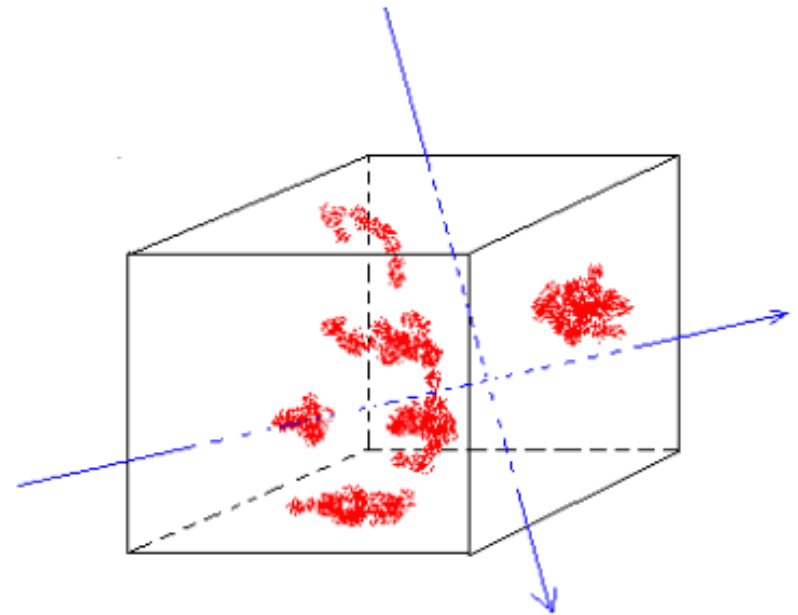
- atgggtaaaataattggtatcgacct

random

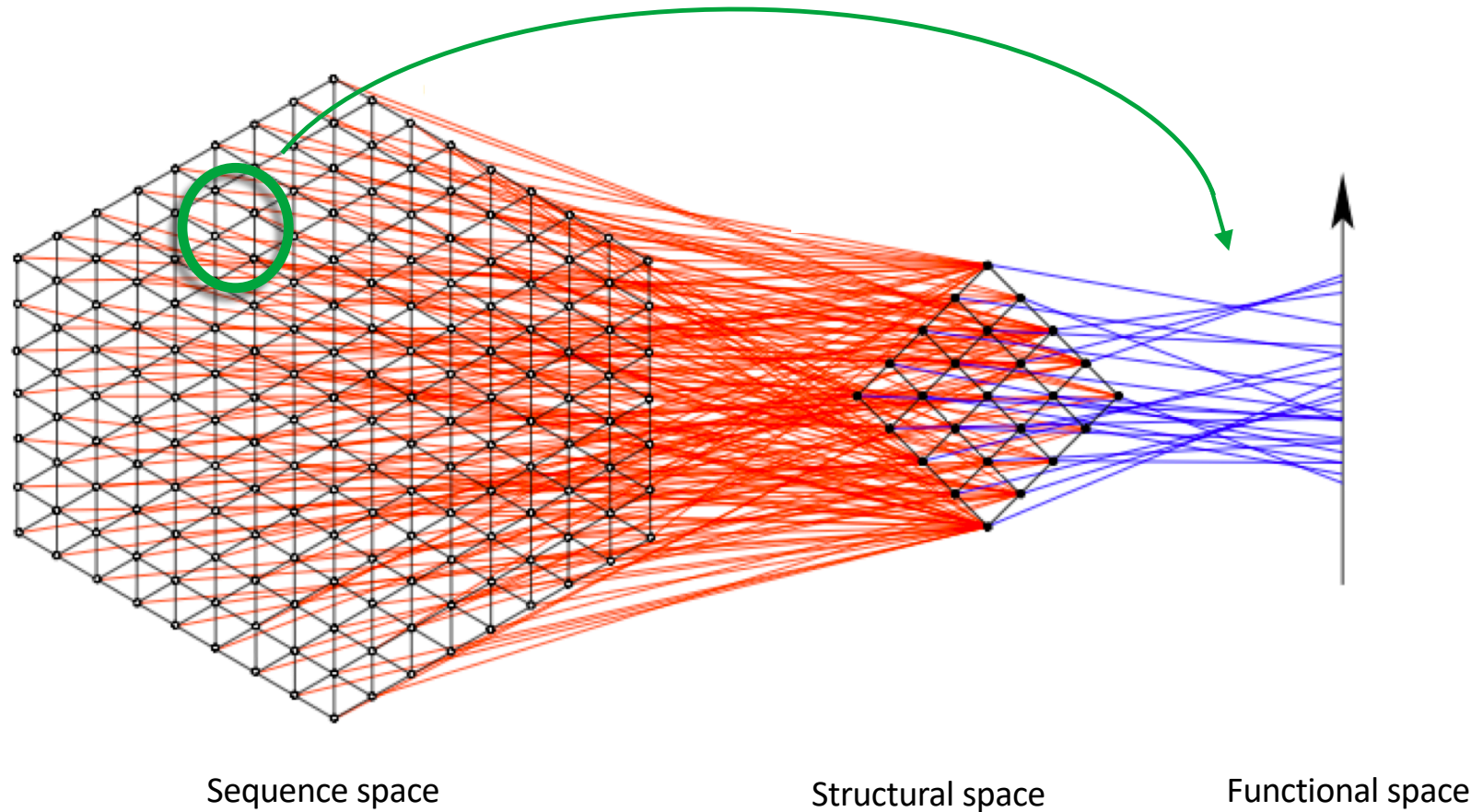
- tgaggtcgtagagtagacca

- 4^n possible sequences

- Real sequences occupy a tiny fraction of all possible sequences



Mapping the spaces: starting from raw seqs

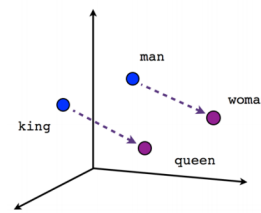


Embedding for categorical variables

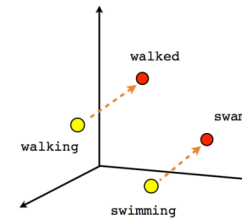
puppy	[1, 0, 0, 0]
dog	[0, 1, 0, 0]
kitten	[0, 0, 1, 0]
cat	[0, 0, 0, 1]



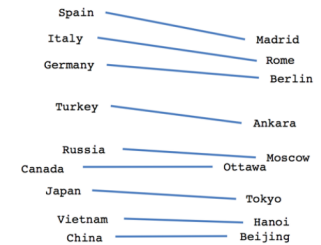
puppy	[0.9, 1.0, 0.0, 0.2]
dog	[1.0, 0.2, 0.0, 0.9]
kitten	[0.0, 1.0, 0.5, 0.1]
cat	[0.0, 0.2, 1.0, 1.0]



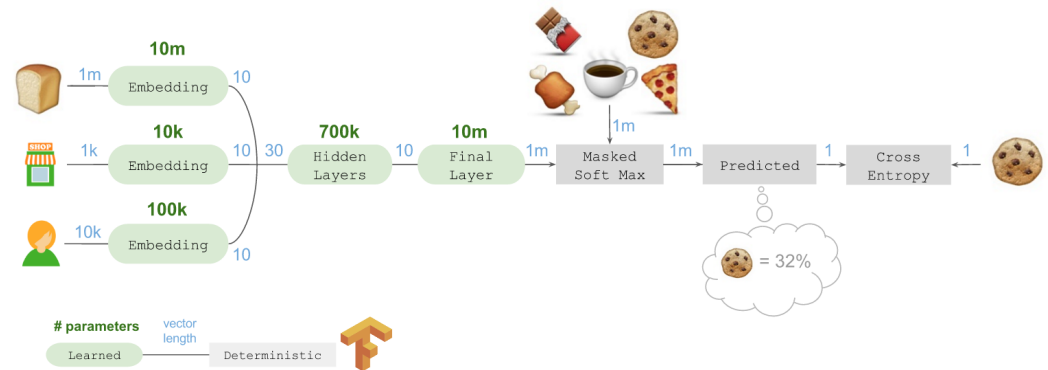
Male-Female



Verb tense

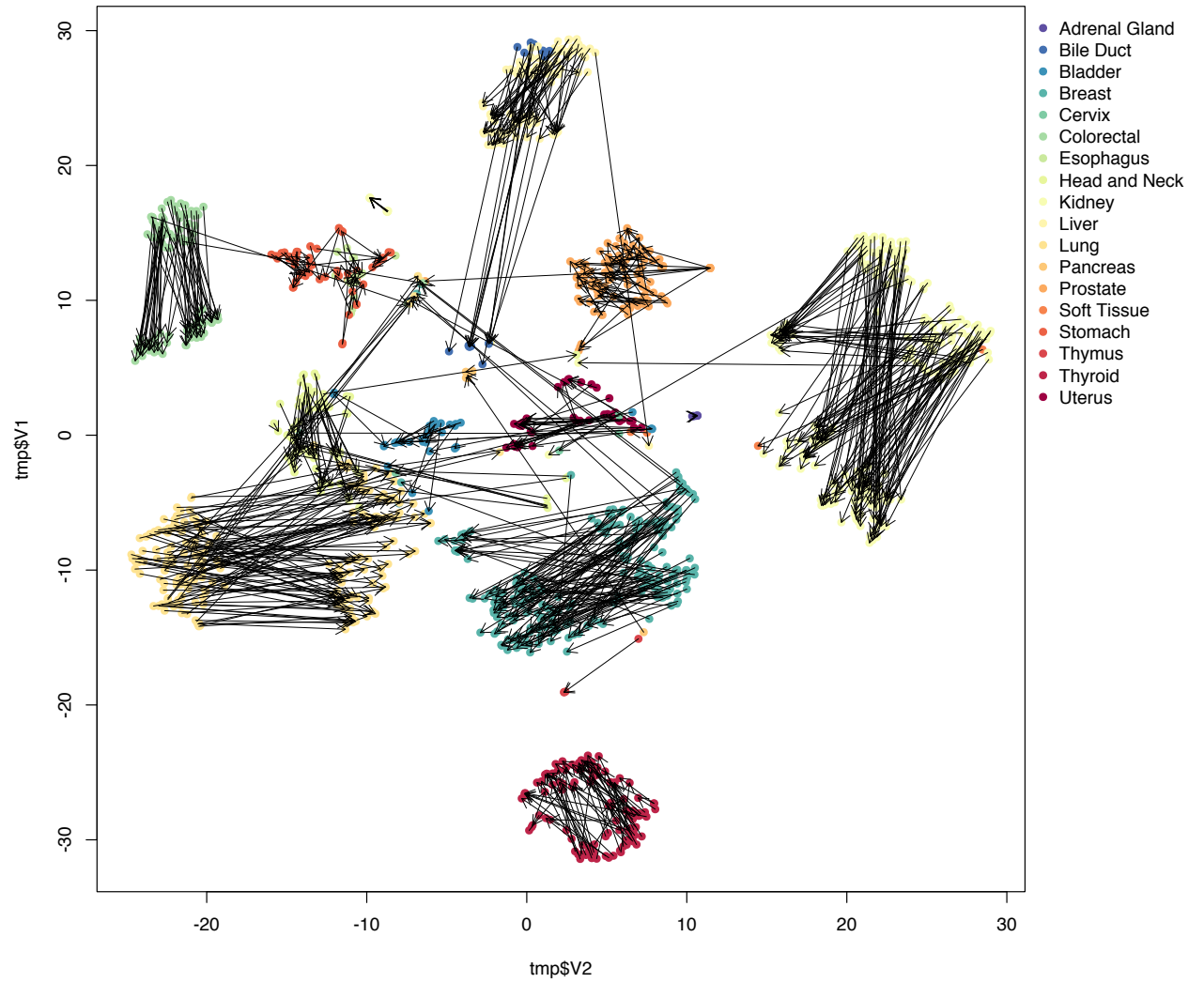


Country-Capital



From the Instacart blog post 'Deep Learning with Emojis (not Math)'

Normal => Tumor



BIOLOGICAL STRUCTURE AND FUNCTION EMERGE FROM SCALING UNSUPERVISED LEARNING TO 250 MILLION PROTEIN SEQUENCES

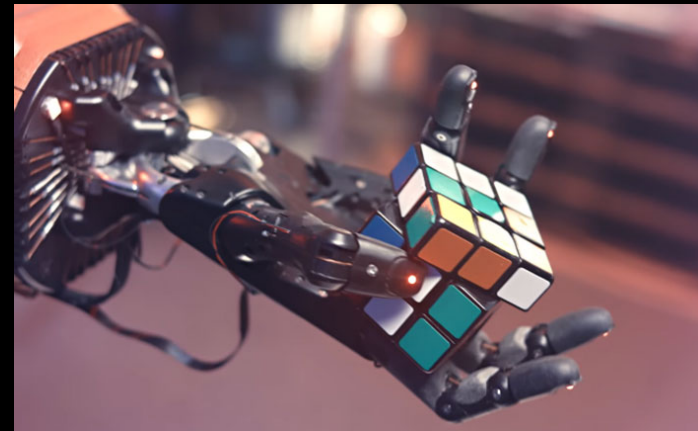
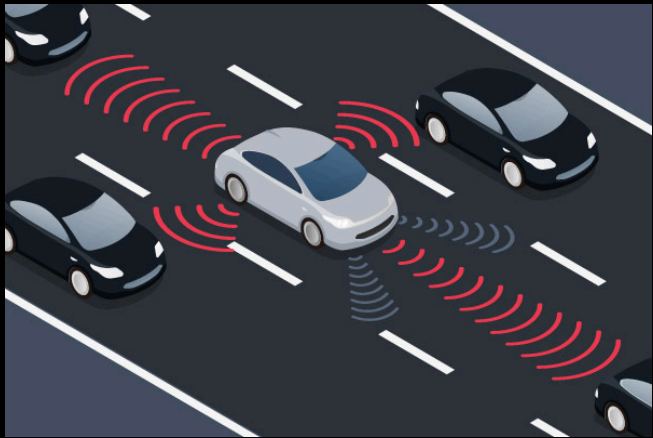
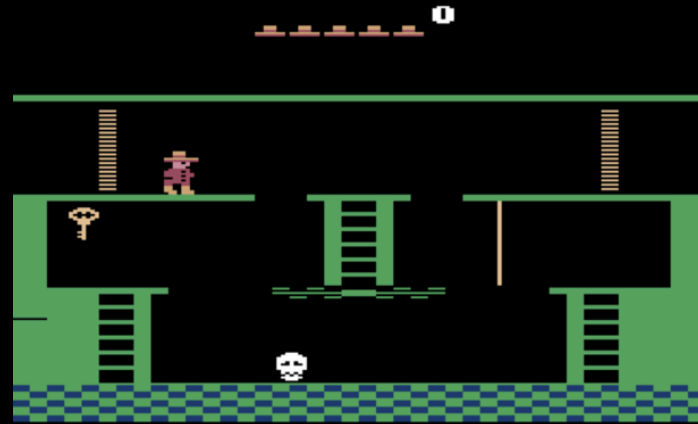
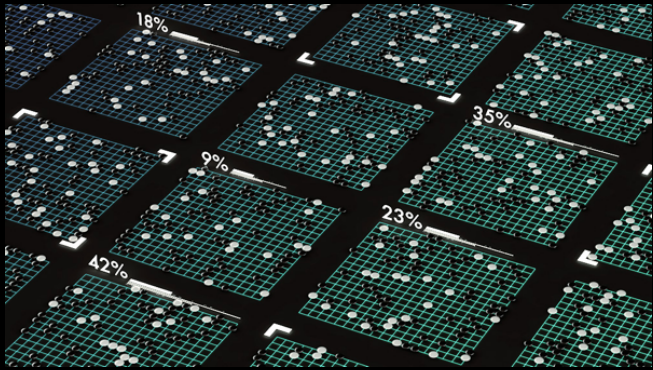
Alexander Rives ^{*†‡} **Siddharth Goyal** ^{*§} **Joshua Meier** ^{*§} **Demi Guo** ^{*§}
Myle Ott [§] **C. Lawrence Zitnick** [§] **Jerry Ma** ^{†§} **Rob Fergus** ^{†‡§}

In the field of artificial intelligence, a combination of scale in data and model capacity enabled by unsupervised learning has led to major advances in representation learning and statistical generation. In biology, the anticipated growth of sequencing promises unprecedented data on natural sequence diversity. Learning the natural distribution of evolutionary protein sequence variation is a logical step toward predictive and generative modeling for biology. [To this end we use unsupervised learning to train a deep contextual language model on 86 billion amino acids across 250 million sequences spanning evolutionary diversity.](#) The resulting model maps raw sequences to representations of biological properties without labels or prior domain knowledge. The learned representation space organizes sequences at multiple levels of biological granularity from the biochemical to proteomic levels. Learning recovers information about protein structure: secondary structure and residue-residue contacts can be extracted by linear projections from learned representations. With small amounts of labeled data, the ability to identify tertiary contacts is further improved. Learning on full sequence diversity rather than individual protein families increases recoverable information about secondary structure. We show the networks generalize by adapting them to variant activity prediction from sequences only, with results that are comparable to a state-of-the-art variant predictor that uses evolutionary and structurally derived features.

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[‡]Dept. of Computer Science, New York University, USA

[§]Facebook AI Research, USA



What is Machine Learning?

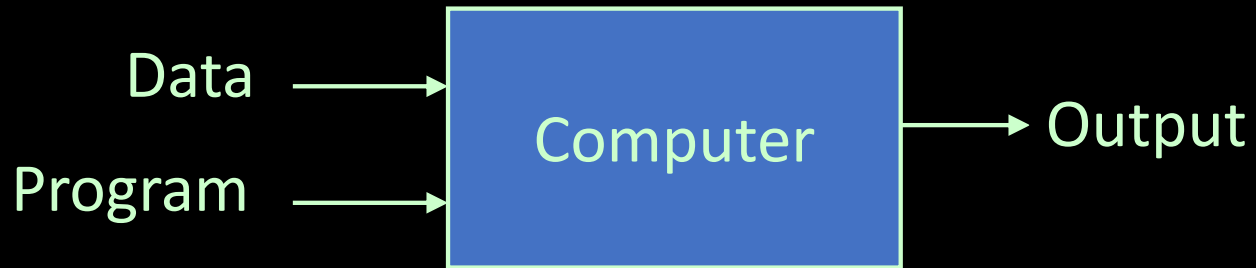
The complexity in traditional computer programming is in the code (programs that people write). In machine learning, algorithms (programs) are in principle simple and the complexity (structure) is in the data. Is there a way that we can automatically learn that structure? That is what is at the heart of machine learning.

-- Andrew Ng

That is, machine learning is the about the construction and study of systems that can learn from data. This is very different than traditional computer programming.

The Cartoon Form

Traditional Programming



Machine Learning

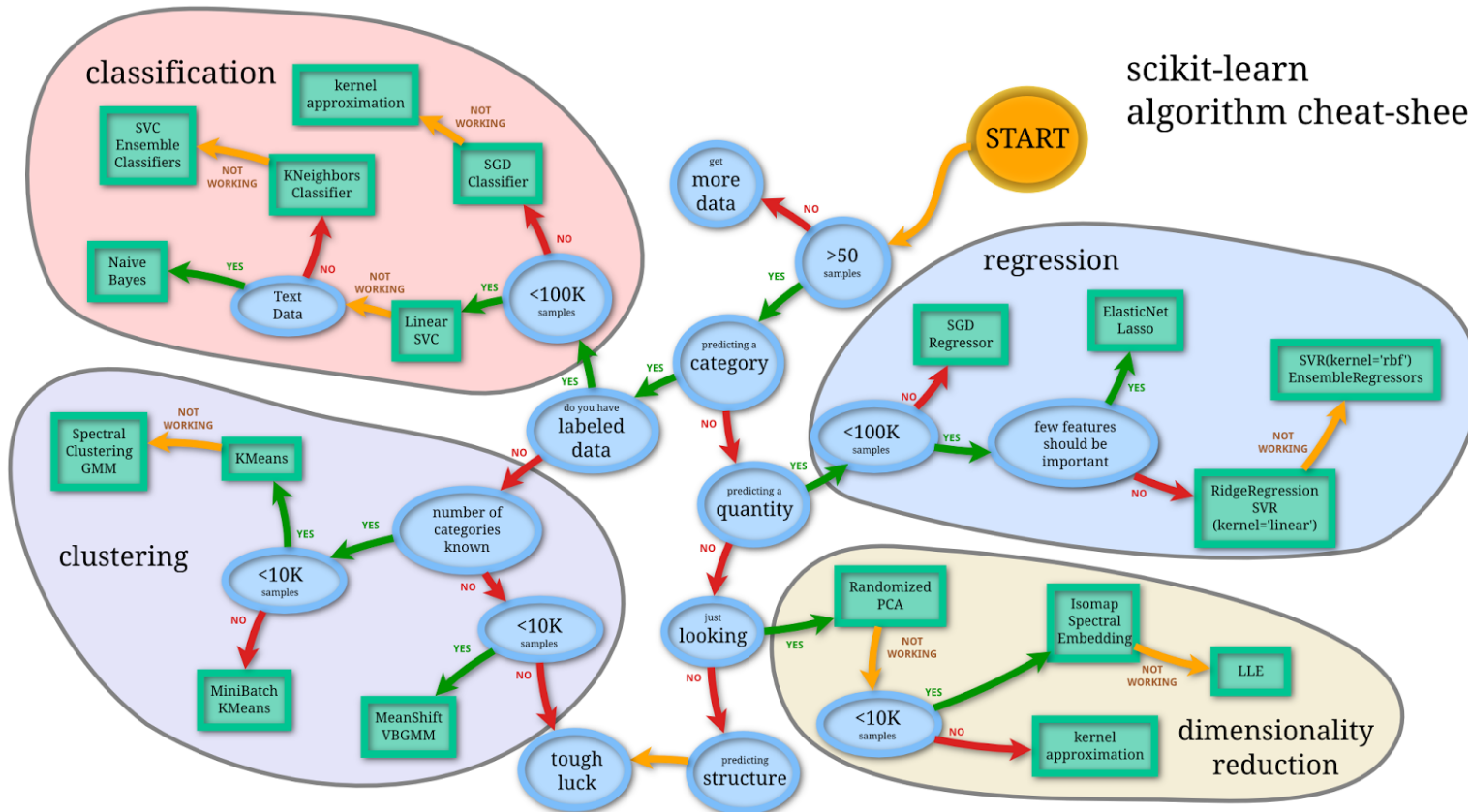


Examples in applying ML to NGS data

- Classifying cancer type with gene expression profiles
- Removing study bias in tumor gene expression profiles
- Classifying cancer type with SNP data
- Drug response prediction - introduction

Four Typical Problems

scikit-learn
algorithm cheat-sheet



Deep learning in biology and medicine

<https://github.com/greenelab/deep-review>

Keras.js

Tensorflow playground



Epoch
000,564

Learning rate
0.03

Activation
Tanh

Regularization
None

Regularization rate
0

Problem type
Classification

DATA

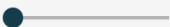
Which dataset do you want to use?



Ratio of training to test data: 50%



Noise: 0



Batch size: 10



REGENERATE

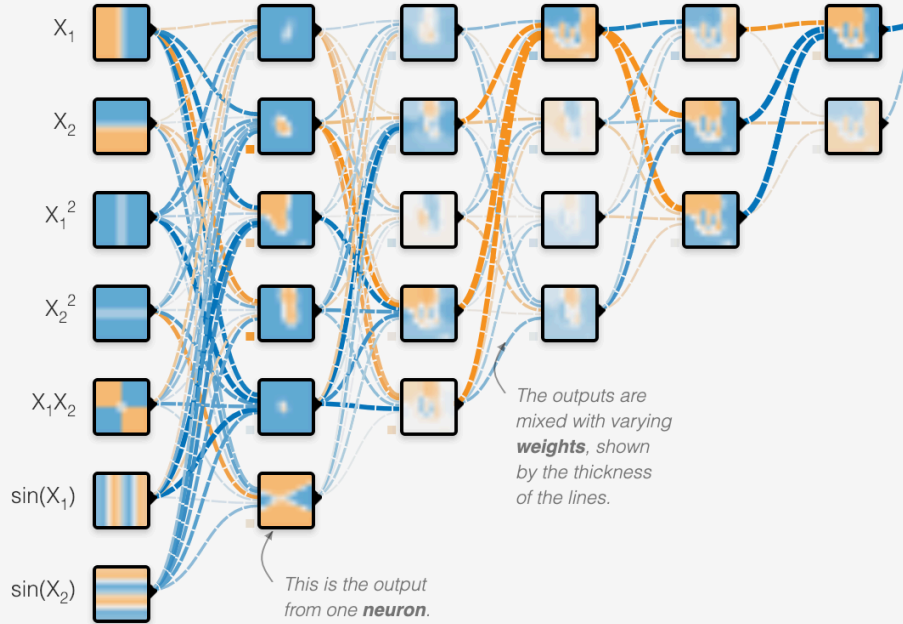
FEATURES

Which properties do you want to feed in?

- X_1
- X_2
- X_1^2
- X_2^2
- $X_1 X_2$
- $\sin(X_1)$
- $\sin(X_2)$

5 HIDDEN LAYERS

6 neurons 5 neurons 4 neurons 3 neurons 2 neurons

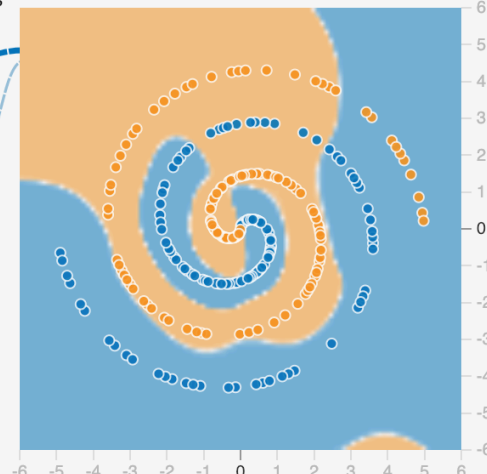
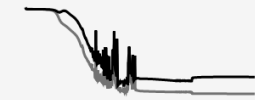


The outputs are mixed with varying weights, shown by the thickness of the lines.

This is the output from one neuron. Hover to see it larger.

OUTPUT

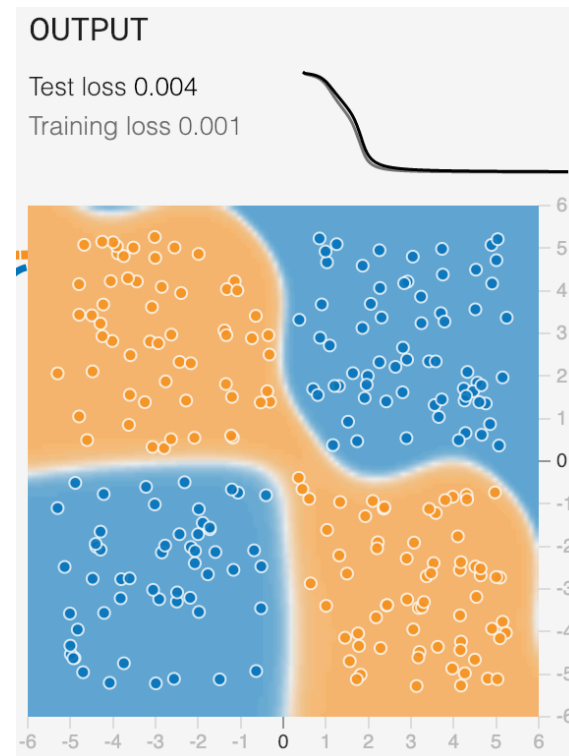
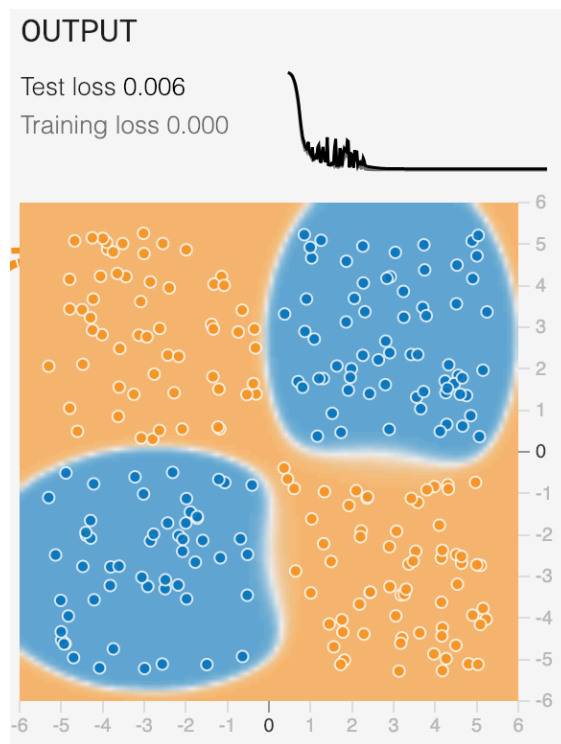
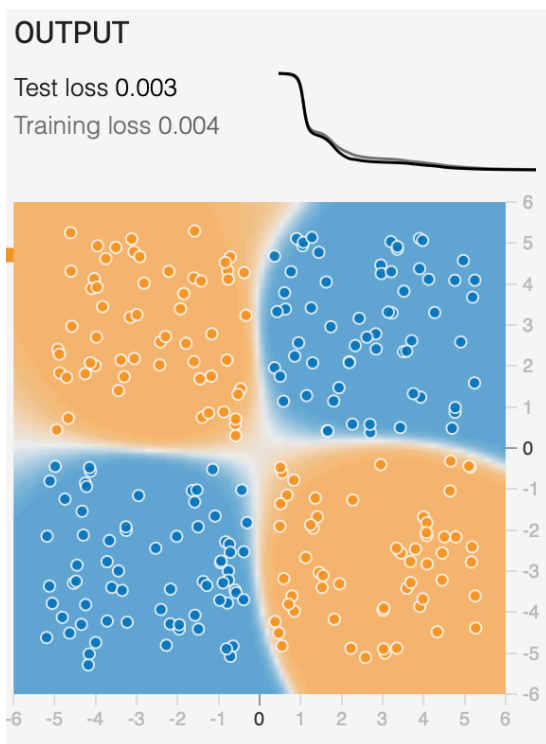
Test loss 0.223
Training loss 0.154



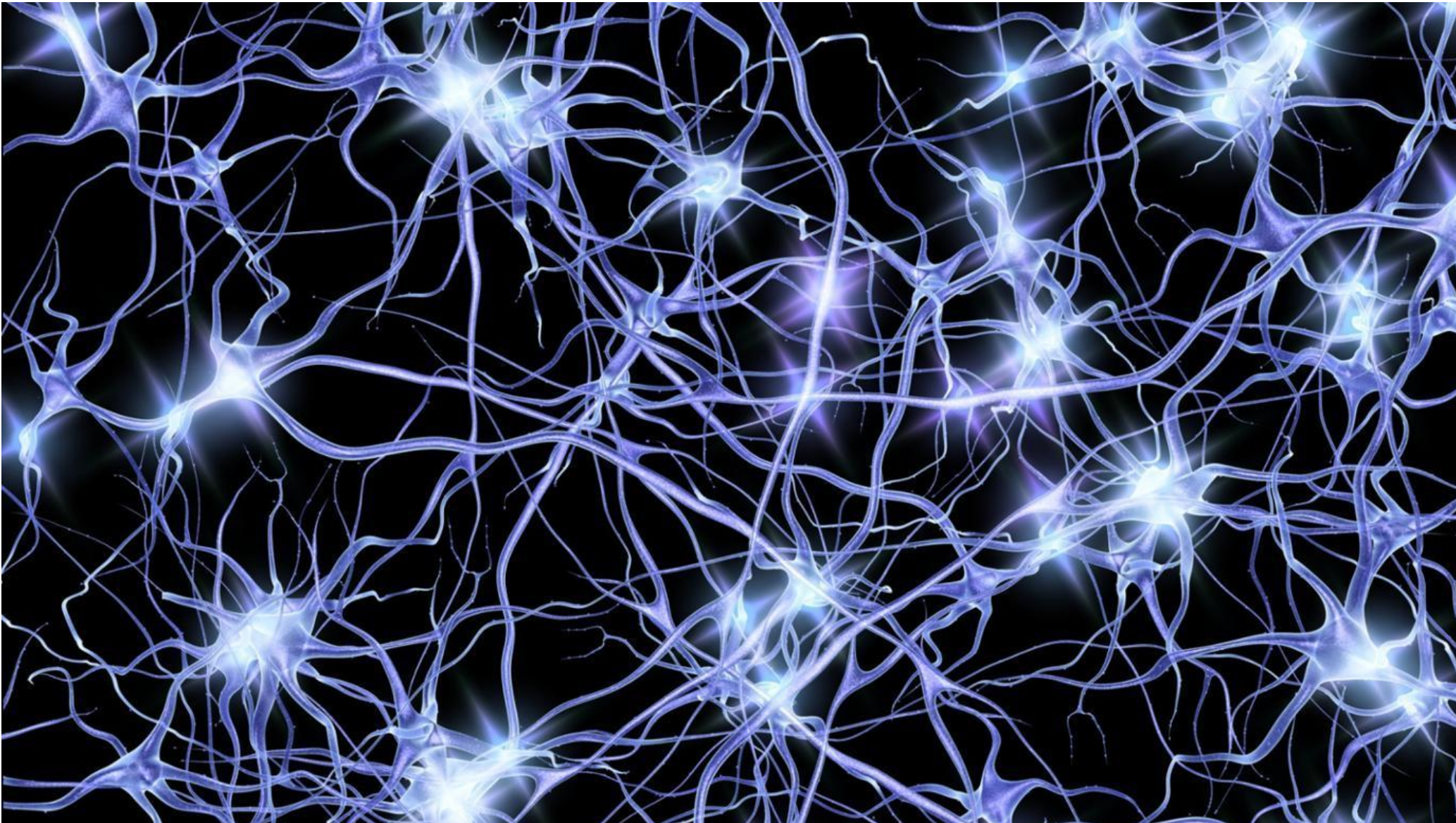
Colors shows data, neuron and weight values. -1 0 1

Show test data Discretize output

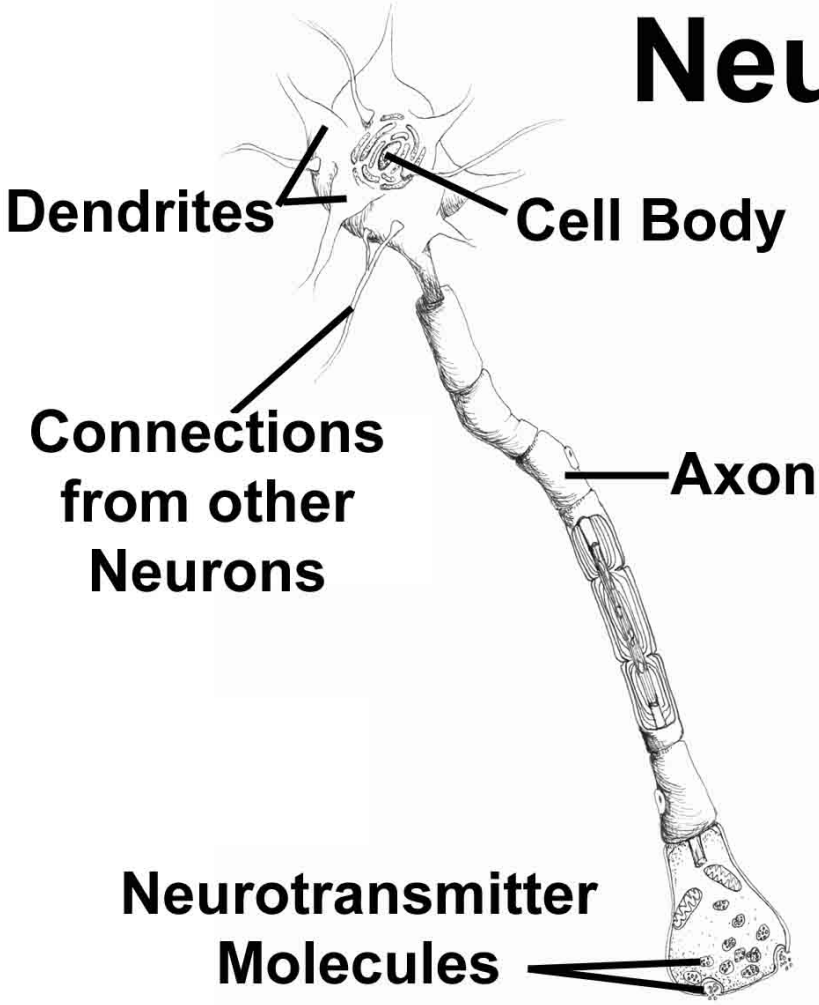
A few “good” runs



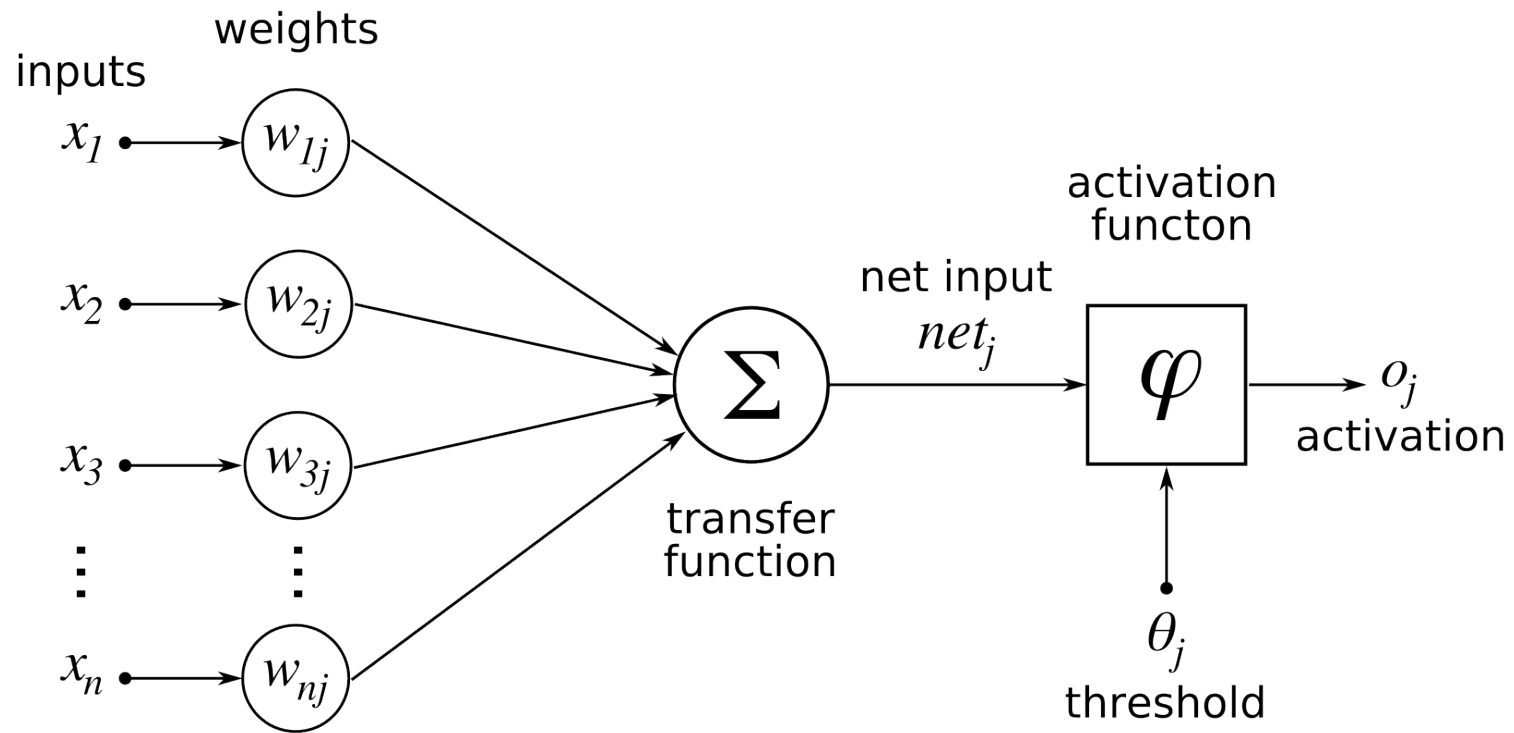
Deep Learning Basics



Neuron



Mathematical Model of a Neuron

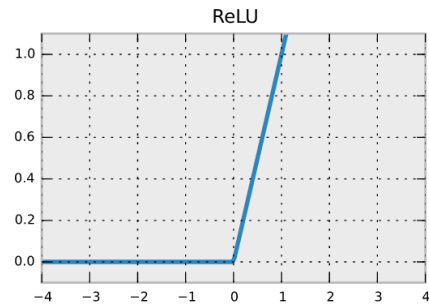
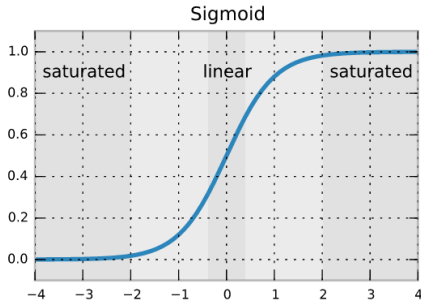


$$z = w \cdot x + b$$

$$o = \varphi(z)$$

Activation

$$\sigma(z) \equiv \frac{1}{1 + e^{-z}}$$



Name	Plot	Equation	Derivative
Identity		$f(x) = x$	$f'(x) = 1$
Binary step		$f(x) = \begin{cases} 0 & \text{for } x < 0 \\ 1 & \text{for } x \geq 0 \end{cases}$	$f'(x) = \begin{cases} 0 & \text{for } x \neq 0 \\ ? & \text{for } x = 0 \end{cases}$
Logistic (a.k.a Soft step)		$f(x) = \frac{1}{1 + e^{-x}}$	$f'(x) = f(x)(1 - f(x))$
TanH		$f(x) = \tanh(x) = \frac{2}{1 + e^{-2x}} - 1$	$f'(x) = 1 - f(x)^2$
ArcTan		$f(x) = \tan^{-1}(x)$	$f'(x) = \frac{1}{x^2 + 1}$
Rectified Linear Unit (ReLU)		$f(x) = \begin{cases} 0 & \text{for } x < 0 \\ x & \text{for } x \geq 0 \end{cases}$	$f'(x) = \begin{cases} 0 & \text{for } x < 0 \\ 1 & \text{for } x \geq 0 \end{cases}$
Parameteric Rectified Linear Unit (PReLU) ^[2]		$f(x) = \begin{cases} \alpha x & \text{for } x < 0 \\ x & \text{for } x \geq 0 \end{cases}$	$f'(x) = \begin{cases} \alpha & \text{for } x < 0 \\ 1 & \text{for } x \geq 0 \end{cases}$
Exponential Linear Unit (ELU) ^[3]		$f(x) = \begin{cases} \alpha(e^x - 1) & \text{for } x < 0 \\ x & \text{for } x \geq 0 \end{cases}$	$f'(x) = \begin{cases} f(x) + \alpha & \text{for } x < 0 \\ 1 & \text{for } x \geq 0 \end{cases}$
SoftPlus		$f(x) = \log_e(1 + e^x)$	$f'(x) = \frac{1}{1 + e^{-x}}$

Loss and Accuracy

Regression problem

y_true: [-50, -10, +80]

y_pred: [-30, +10, +60]

MSE = 0.04

Classification *view*

[1, 1, 0]

[1, 0, 1]

ACC = 0.67

y_true: [-50, -10, +80]

y_pred: [-50, -10, +45]

MSE = 0.04

[1, 1, 0]

ACC = 1.00

Classification loss

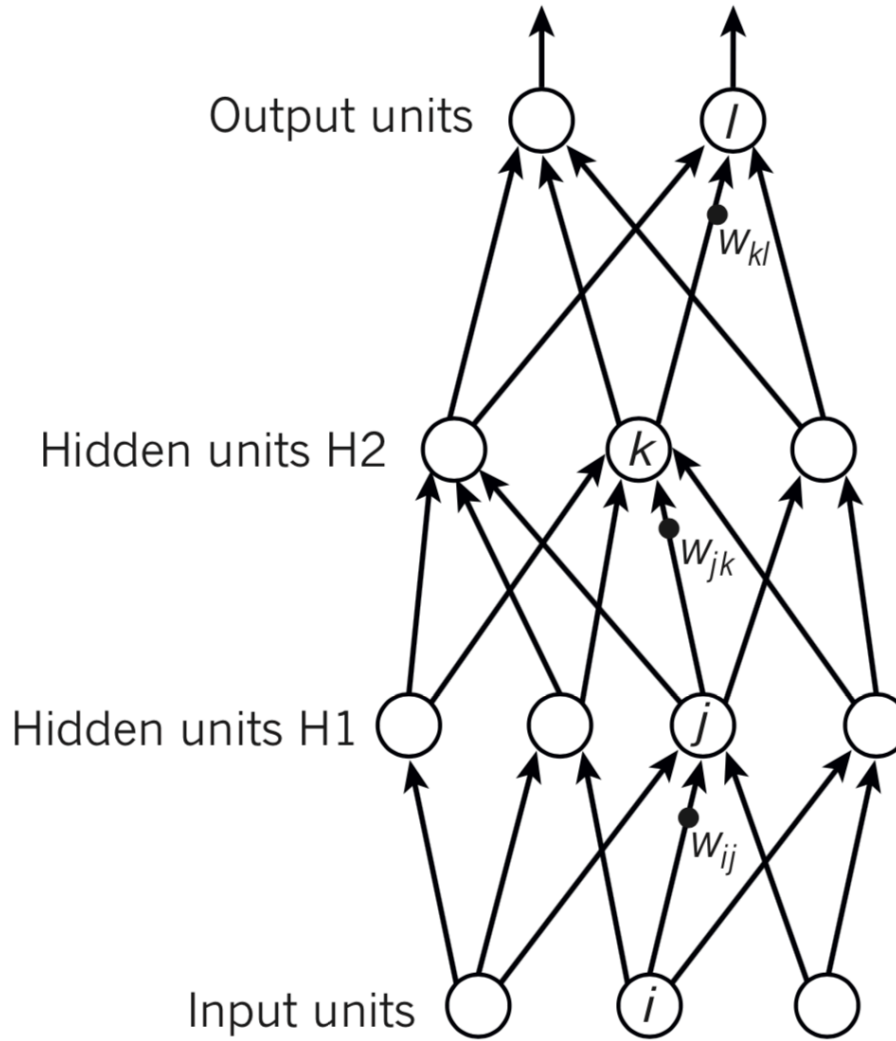
- Softmax
- Cross entropy

How different are the two probability distributions?

scores	softmax →	probabilities	true probabilities	cross entropy
0.6	$\frac{e^{z_j}}{\sum_{k=1}^K e^{z_k}}$	0.059	0.1	0.866
-1		0.012	0.2	
3.2		0.797	0.6	
1.4		0.132	0.1	

$$H(p, q) = - \sum_x p(x) \log q(x)$$

c



$$y_l = f(z_l)$$

$$z_l = \sum_{k \in H2} w_{kl} y_k$$

$$y_k = f(z_k)$$

$$z_k = \sum_{j \in H1} w_{jk} y_j$$

$$y_j = f(z_j)$$

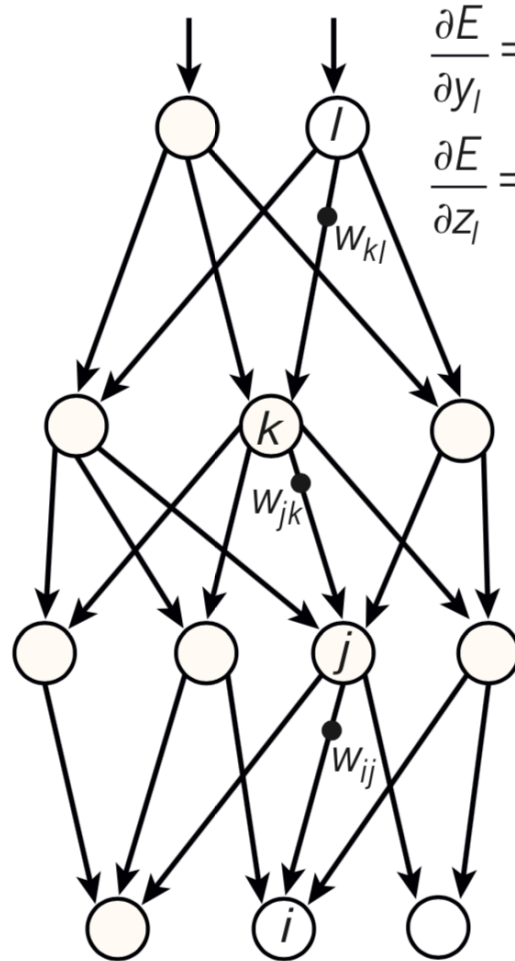
$$z_j = \sum_{i \in \text{Input}} w_{ij} x_i$$

d

Compare outputs with correct answer to get error derivatives

$$\frac{\partial E}{\partial y_k} = \sum_{l \in \text{out}} w_{kl} \frac{\partial E}{\partial z_l}$$

$$\frac{\partial E}{\partial z_k} = \frac{\partial E}{\partial y_k} \frac{\partial y_k}{\partial z_k}$$



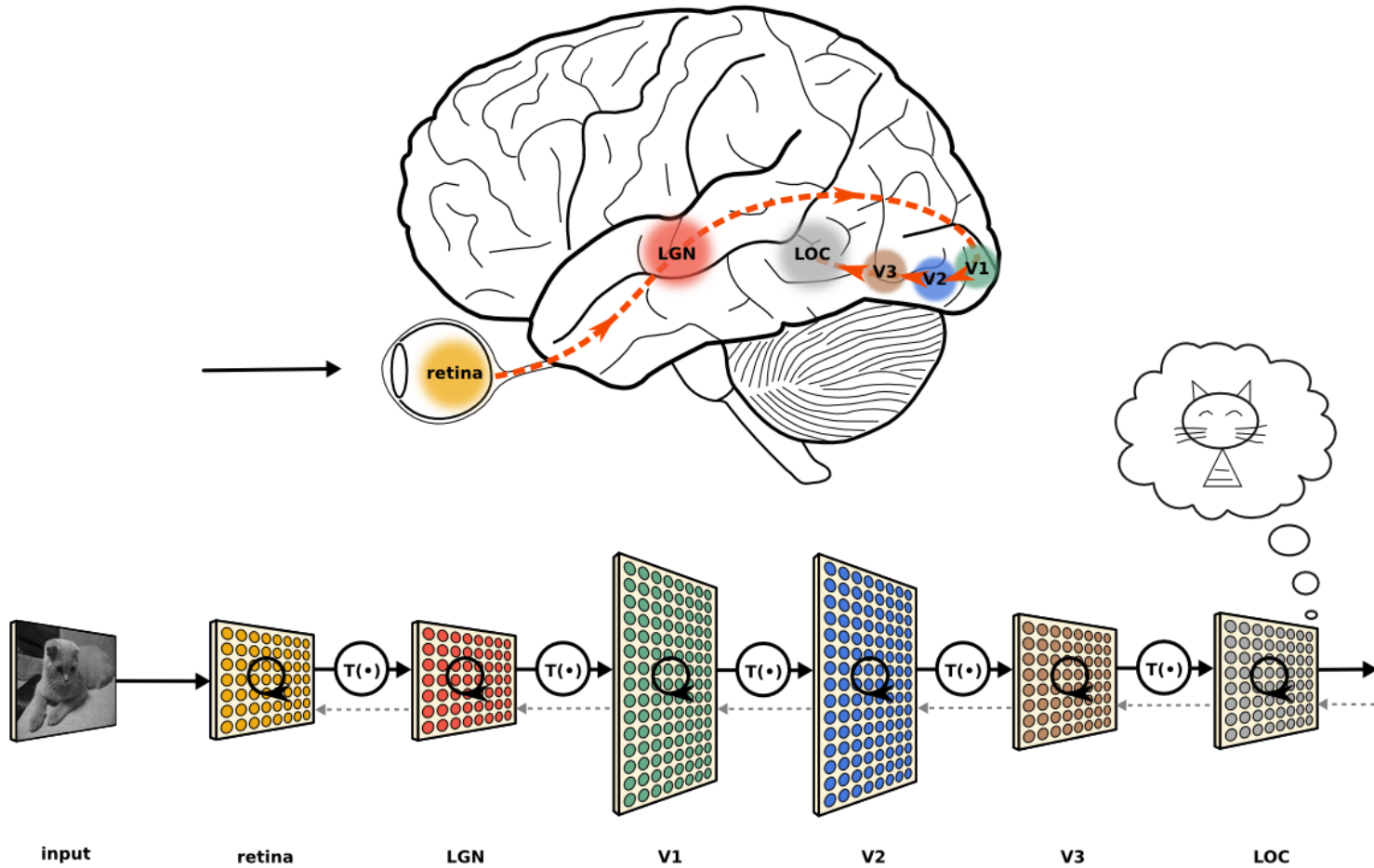
$$\frac{\partial E}{\partial y_l} = y_l - t_l$$

$$\frac{\partial E}{\partial z_l} = \frac{\partial E}{\partial y_l} \frac{\partial y_l}{\partial z_l}$$

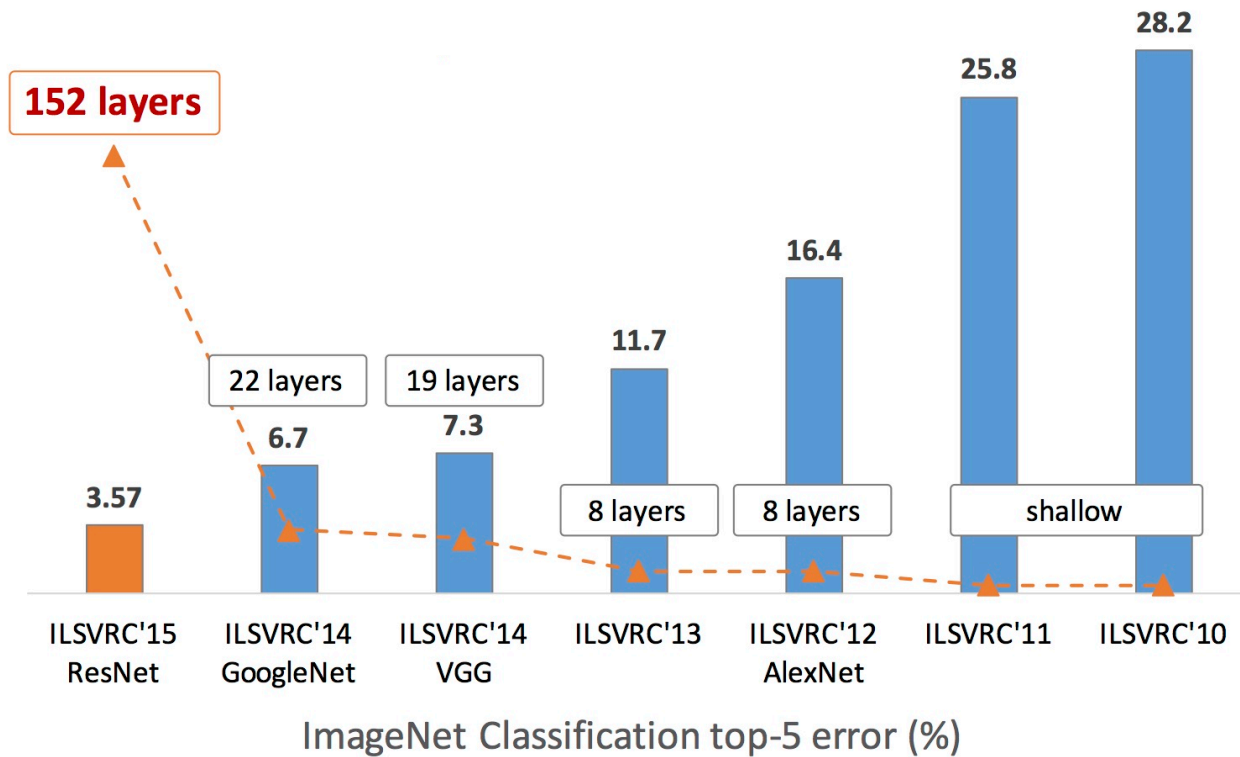
$$\frac{\partial E}{\partial y_j} = \sum_{k \in H2} w_{jk} \frac{\partial E}{\partial z_k}$$

$$\frac{\partial E}{\partial z_j} = \frac{\partial E}{\partial y_j} \frac{\partial y_j}{\partial z_j}$$

Human Vision System

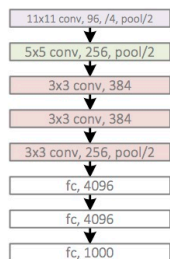


Increasing Depth Works...

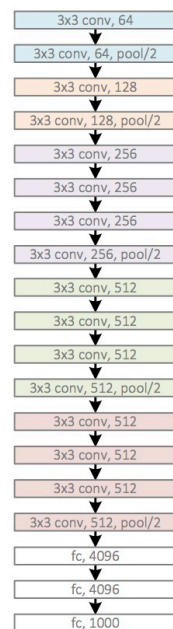


Example CNNs Structures from ILSVRC Winners

AlexNet, 8 layers
(ILSVRC 2012)



VGG, 19 layers
(ILSVRC 2014)



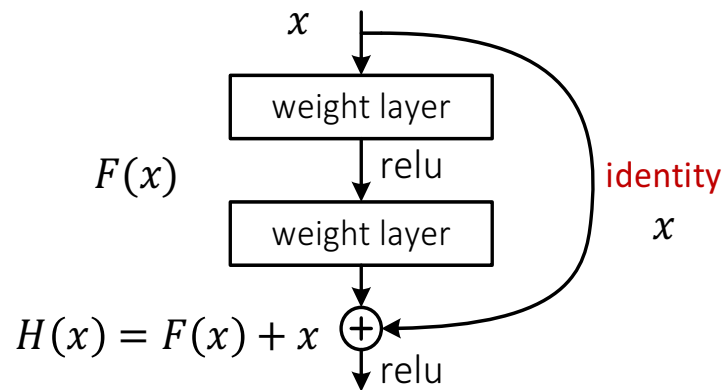
GoogleNet, 22 layers
(ILSVRC 2014)



Kaiming He, Xiangyu Zhang, Shaoqing Ren, & Jian Sun. "Deep Residual Learning for Image Recognition". CVPR 2016.

Deep Residual Learning

- $F(x)$ is a **residual** mapping w.r.t. **identity**

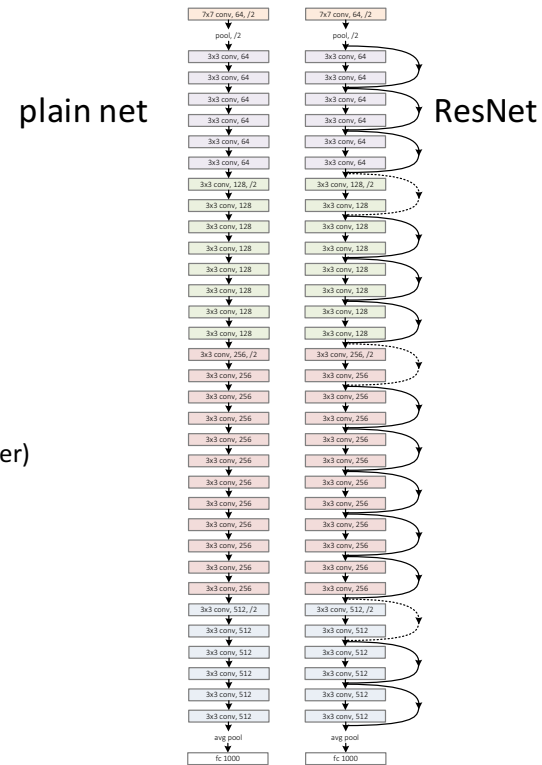


- If identity were optimal, easy to set weights as 0
- If optimal mapping is closer to identity, easier to find small fluctuations

Deep Residual Learning

Network “Design”

- Keep it simple
- Our basic design (VGG-style)
 - all 3x3 conv (almost)
 - spatial size /2 => # filters x2 (~same complexity per layer)
 - **Simple design; just deep!**
- Other remarks:
 - no hidden fc
 - no dropout



Dropout

SRIVASTAVA, HINTON, KRIZHEVSKY, SUTSKEVER AND SALAKHUTDINOV

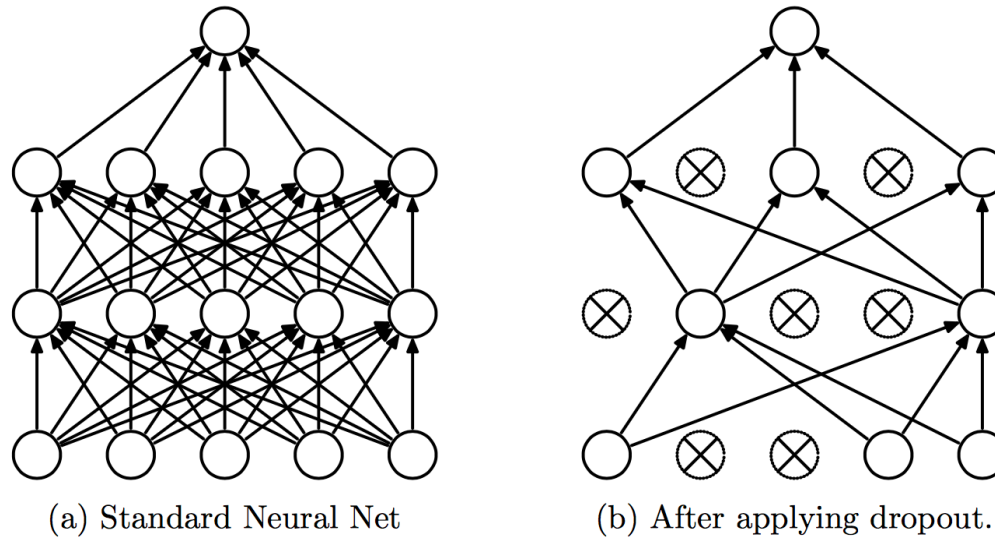


Figure 1: Dropout Neural Net Model. **Left:** A standard neural net with 2 hidden layers. **Right:** An example of a thinned net produced by applying dropout to the network on the left. Crossed units have been dropped.



Andres Torrubia @antor · Oct 18



Replying to @radekosmulski

Sometimes it feels like 🙏

QUESTION 8/10

As demonstrated in research, what is the trick that enables training a 10,000 layer deep neural network?

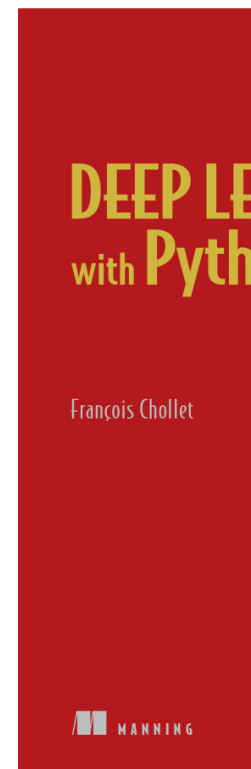
- dropout
- batchnorm
- initialization
- thoughts and prayers

Submit answer



Essential Deep Learning Resources

- **Books**
 - Michael Nielsen's free online book on deep learning
 - Deep Learning textbook (Goodfellow et al, 2016)
 - Deep Learning with Python (Keras book)
- **Online courses**
 - FastAI
 - Stanford CS231n
 - Andrew Ng
- **Python**
 - Python Cookbook, 3rd Edition
- **Other resources**
 - PyTorch tutorials
 - Kaggle kernels / discussions; No free hunch interviews
 - Twitter
 - arxiv-sanity



Improving Models with Domain Knowledge

- **Books**
 - Michael Nielsen's free online book on deep learning
 - Deep Learning textbook (Goodfellow et al, 2016)
 - Deep Learning with Python (Keras book)
- **Online courses**
 - FastAI
 - Stanford CS231n
 - Andrew Ng
- **Python**
 - Python Cookbook, 3rd Edition
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 - Twitter
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About 10,000 deep learning papers have been written about "hard-coding priors about a specific task into a NN architecture works better than a lack of prior" -- but they're typically being passed as "architecture XYZ offers superior performance for [overly generic task category]"

749 Likes	168 Retweets
Jun 1, 2019 at 5:19 PM	via Twitter Web Client

← ↻ ❤️ ↗ ⚙️



François Chollet @fchollet 36d

You can always "buy" performance by either training on more data, better data, or by injecting task information into the architecture or the preprocessing. However, this isn't informative about the generalization power of the techniques used (which is the only thing that matters)

*Cancer Type Classification
with RNAseq*

Harmonized Cancer Datasets Genomic Data Commons Data Portal

Get Started by Exploring:



Projects



Data

Perform Advanced Search Queries, such as:

Cases of kidney cancer diagnosed at the age of 20 and below

736 Cases

1,519 Files

CNV data of female brain cancer cases

459 Cases

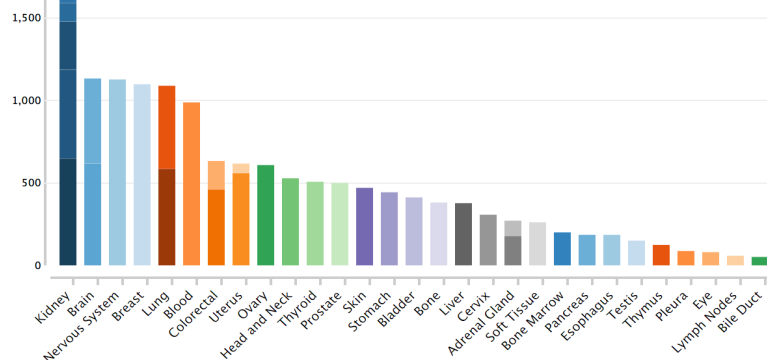
1,788 Files

Gene expression quantification data in TCGA-GBM project

166 Cases

522 Files

Cases by Primary Site



DATA PORTAL SUMMARY

Data Release 6.0 - May 9, 2017

PROJECTS
39

PRIMARY SITE
29

CASES
14,551

FILES
274,724

Infrastructure

Data is continuously being processed and harmonized by the GDC.

View GDC system statistics:

Compute Infrastructure	12,800 Cores	87.96 TB RAM
Storage Infrastructure	4.98 PB Used	5.42 PB Total

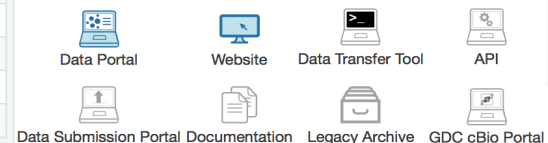
Documentation

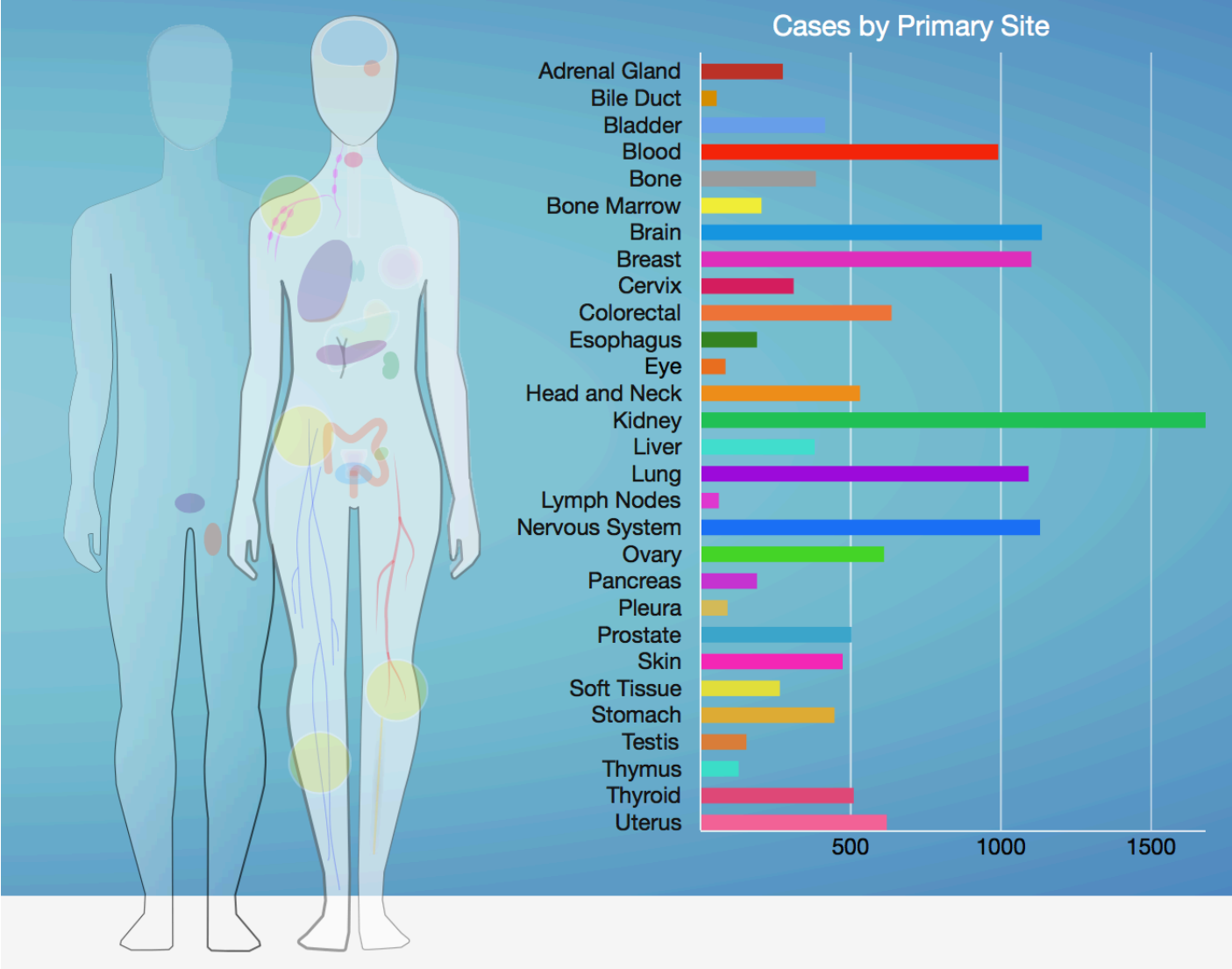
Learn how to use the GDC Data Portal to its full potential with common topics such as:

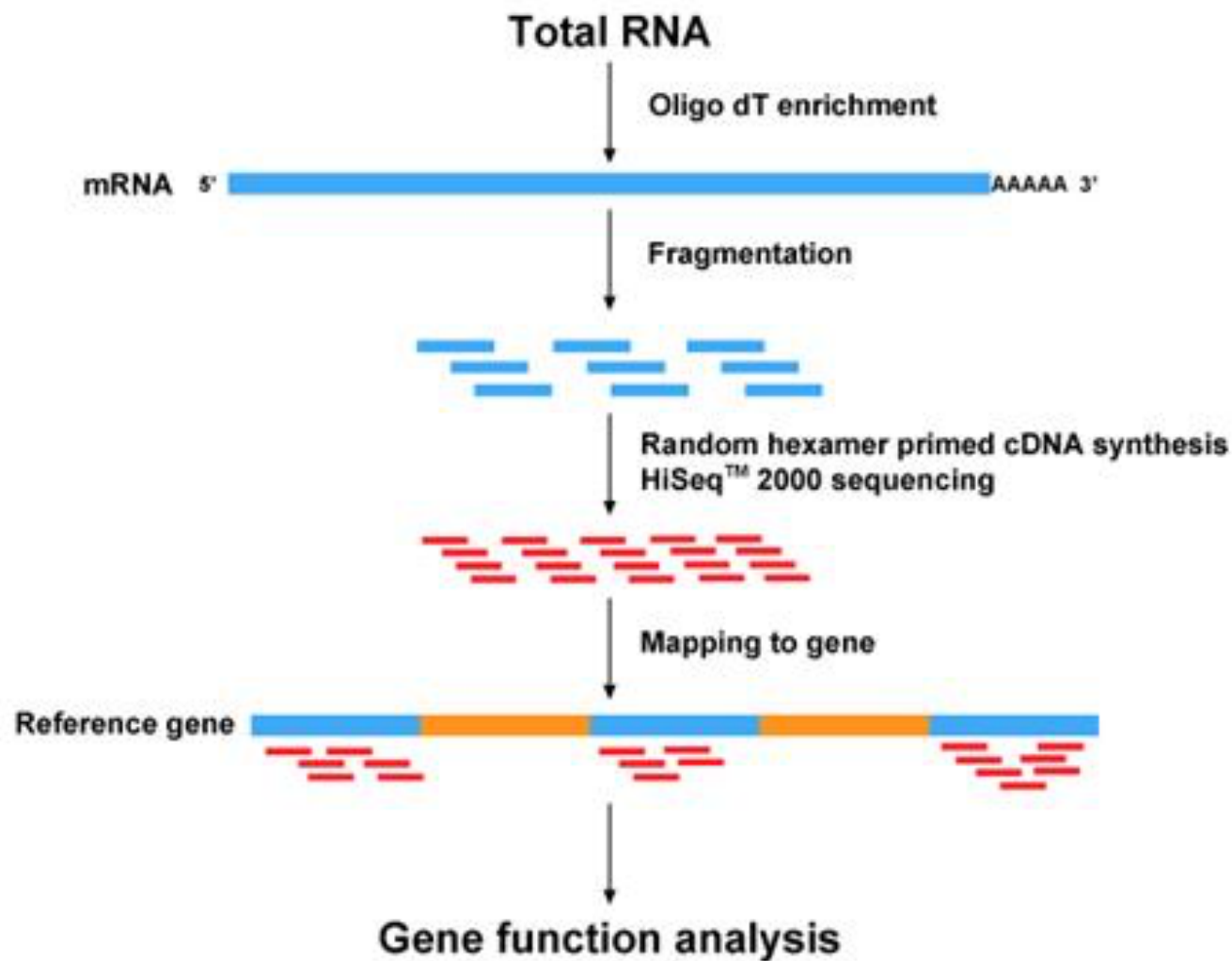
- [Browse Data using Facet Search](#)
- [Search Data with Advanced Search Technology](#)
- [Project Based Data Availability](#)
- [Controlled Access Data](#)
- [Visit the Documentation Website »](#)

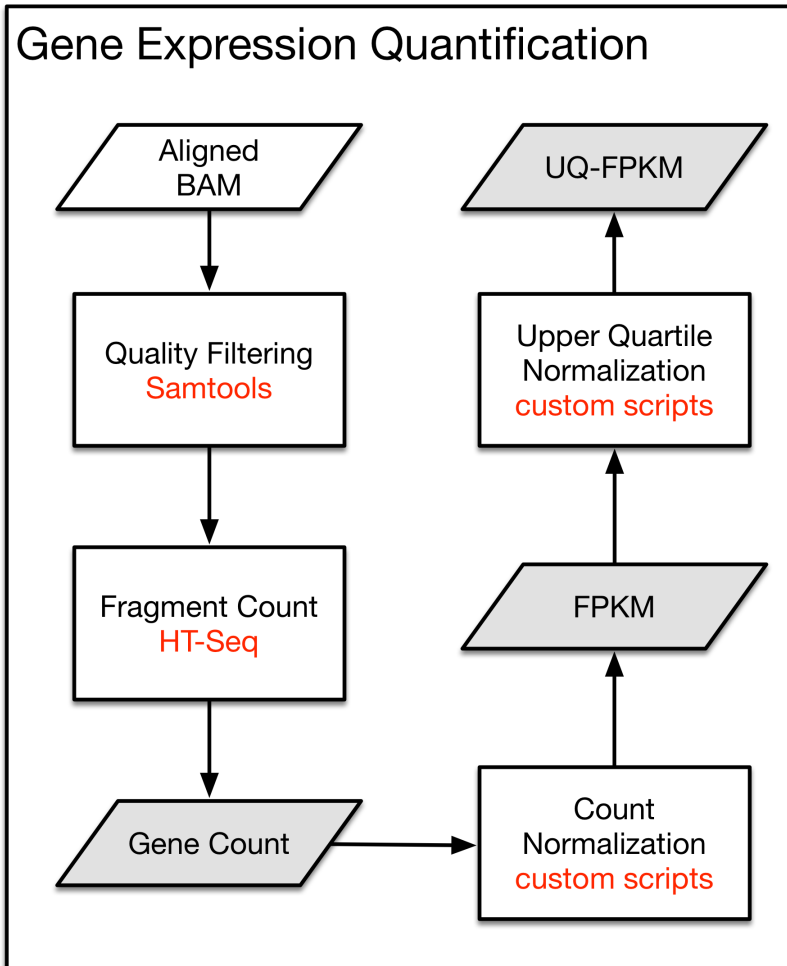
GDC Applications

The GDC Data Portal is a robust data-driven platform that allows cancer researchers and bioinformaticians to search and download cancer data for analysis. The GDC applications include:









RPKM (reads per kilobase per million mapped reads)
Upper Quartile (UQ)

FPKM

The Fragments per Kilobase of transcript per Million mapped reads (FPKM) calculation normalizes read count by dividing it by the gene length and the total number of reads mapped to protein-coding genes.

Upper Quartile FPKM

The upper quartile FPKM (FPKM-UQ) is a modified FPKM calculation in which the total protein-coding read count is replaced by the 75th percentile read count value for the sample.

Calculations

$$FPKM = \frac{RC_g * 10^9}{RC_{pc} * L} \quad FPKM - UQ = \frac{RC_g * 10^9}{RC_{g75} * L}$$

- RC_g : Number of reads mapped to the gene
- RC_{pc} : Number of reads mapped to all protein-coding genes
- RC_{g75} : The 75th percentile read count value for genes in the sample
- L : Length of the gene in base pairs

Note: The read count is multiplied by a scalar (10^9) during normalization to account for the kilobase and 'million mapped reads' units.


Encode numeric columns

- Log transformation: $\log(1+x)$
- Standard scalar: z score
- Rank => Gaussian
- Discretization
- Mark missing values

One Hot Encoding of Categories

State	Binary	One-Hot	Hamming 2	Hamming 3
S0	000	00000001	0000	000000
S1	001	00000010	0011	000111
S2	010	00000100	0101	011001
S3	011	00001000	0110	011110
S4	100	00010000	1001	101010
S5	101	00100000	1010	101101
S6	110	01000000	1100	110011
S7	111	10000000	1111	110100

Open Source Framework Comparison

	Languages	Tutorials and training materials	CNN modeling capability	RNN modeling capability	Architecture: easy-to-use and modular front end	Speed	Multiple GPU support	Keras compatible
Theano	Python, C++	++	++	++	+	++	+	+
TensorFlow	Python	+++	+++	++	+++	++	++	+
Torch	Lua, Python (new)	+	+++	++	++	+++	++	
Caffe	C++	+	++		+	+	+	
MXNet	R, Python, Julia, Scala	++	++	+	++	++	+++	+
Neon	Python	+	++	+	+	++	+	
CNTK	C++	+	+	+++	+	++	+	+

Keras

- <https://keras.io/>
- Minimalist, highly modular neural networks library
- Written in Python
- Capable of running on top of either TensorFlow/Theano and CNTK
- Developed with a focus on enabling fast experimentation



```
from keras.layers import Input, Dense
from keras.models import Model

input_layer = Input(shape=(1000,))
fc_1 = Dense(512, activation='relu')(input_layer)
fc_2 = Dense(256, activation='relu')(fc_1)
output_layer = Dense(10, activation='softmax')(fc_2)

model = Model(input=input_layer, output=output_layer)
model.compile(optimizer='rmsprop',
              loss='categorical_crossentropy',
              metrics=['accuracy'])

model.fit(bow, newsgroups.target)
predictions = model.predict(features).argmax(axis=1)
```

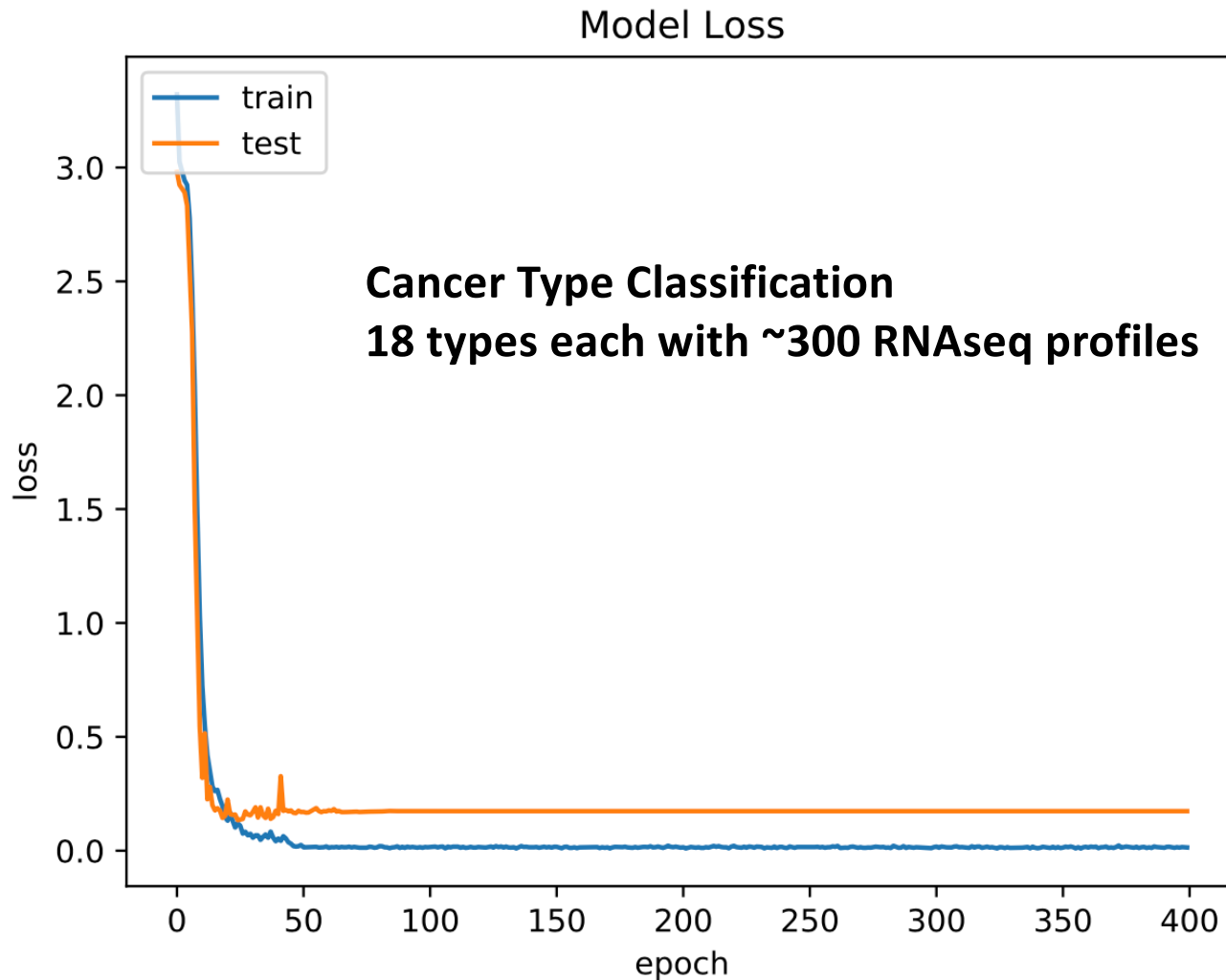
DNN hyperparameter examples

- Data preprocess
 - Positive features: logarithmic transformation $y = \log(1+x)$
 - Mixed features: standard scaler
- Number of hidden layers: 4
- Number of neurons in hidden layers: 4000-2000-1000-1000
- Activation function: ReLU
- Dropouts: Input: 0%; layers 1,2,3: 25%; layer 4: 10%
- Initialization: no unsupervised pretraining
- Optimization: learning rate = 0.05, momentum = 0.9, and weight decay = 0.0001
- Training epochs: as large as possible (dropout can prevent overfitting)

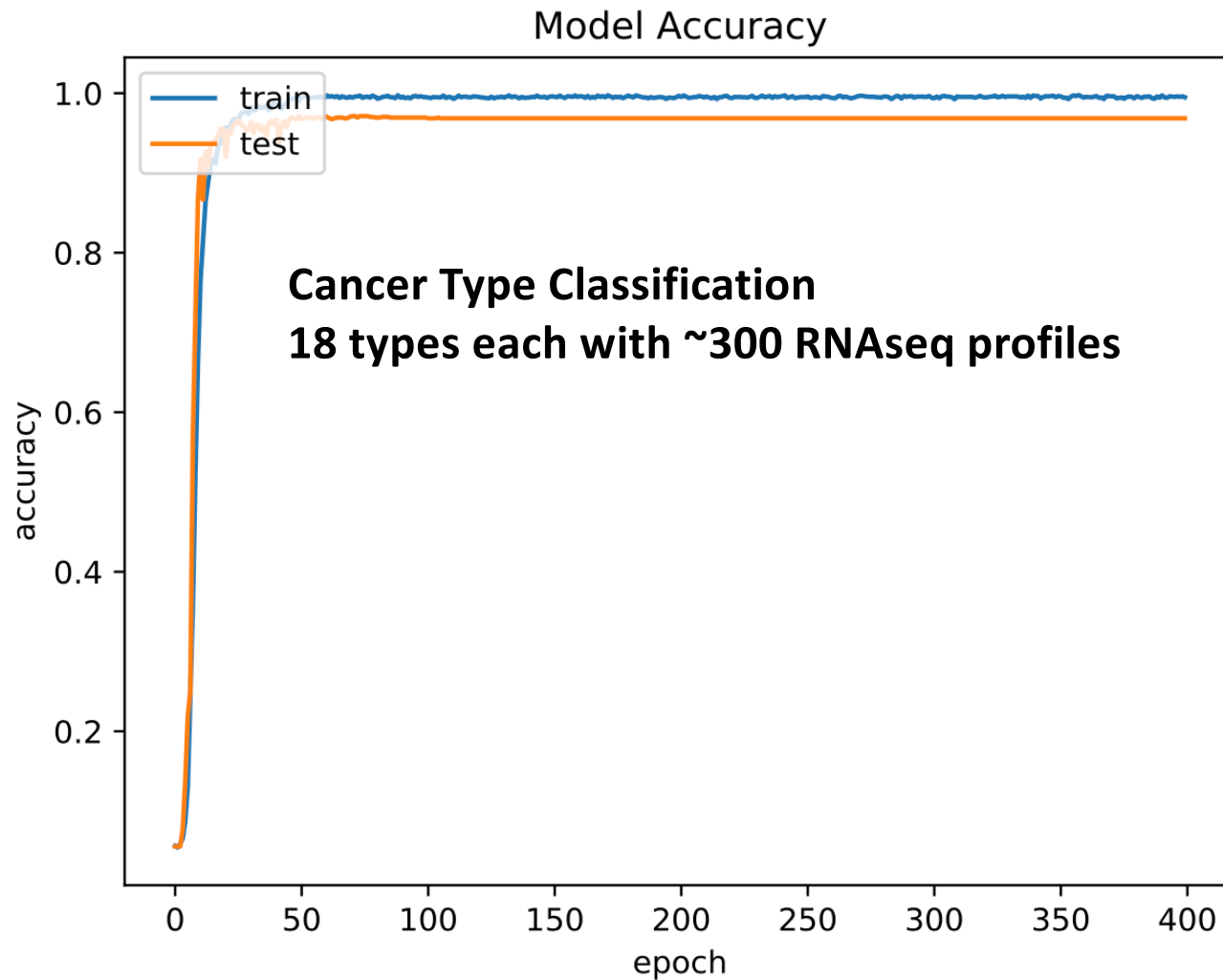
Code Examples

Cancer Type Classification

```
4320/4320 [=====] - 87s - loss: 3.2885 - acc: 0.0537 - val_loss: 2.9542 - val_acc: 0.0556  
Epoch 2/400  
4320/4320 [=====] - 76s - loss: 2.9777 - acc: 0.0752 - val_loss: 2.8273 - val_acc: 0.1083  
Epoch 3/400  
4320/4320 [=====] - 78s - loss: 2.8117 - acc: 0.1176 - val_loss: 2.5971 - val_acc: 0.2194  
Epoch 4/400  
4320/4320 [=====] - 77s - loss: 2.5094 - acc: 0.2060 - val_loss: 2.1191 - val_acc: 0.3306  
Epoch 5/400  
4320/4320 [=====] - 78s - loss: 2.0385 - acc: 0.3442 - val_loss: 1.6411 - val_acc: 0.4648  
Epoch 6/400  
4320/4320 [=====] - 75s - loss: 1.4995 - acc: 0.5079 - val_loss: 0.9846 - val_acc: 0.7704  
Epoch 7/400  
4320/4320 [=====] - 77s - loss: 1.0688 - acc: 0.6481 - val_loss: 0.5628 - val_acc: 0.8796  
Epoch 8/400  
4320/4320 [=====] - 76s - loss: 0.7657 - acc: 0.7461 - val_loss: 0.4952 - val_acc: 0.8509  
Epoch 9/400  
4320/4320 [=====] - 76s - loss: 0.5729 - acc: 0.8123 - val_loss: 0.2803 - val_acc: 0.9287  
Epoch 10/400  
4320/4320 [=====] - 79s - loss: 0.4389 - acc: 0.8620 - val_loss: 0.1962 - val_acc: 0.9398  
Epoch 11/400
```

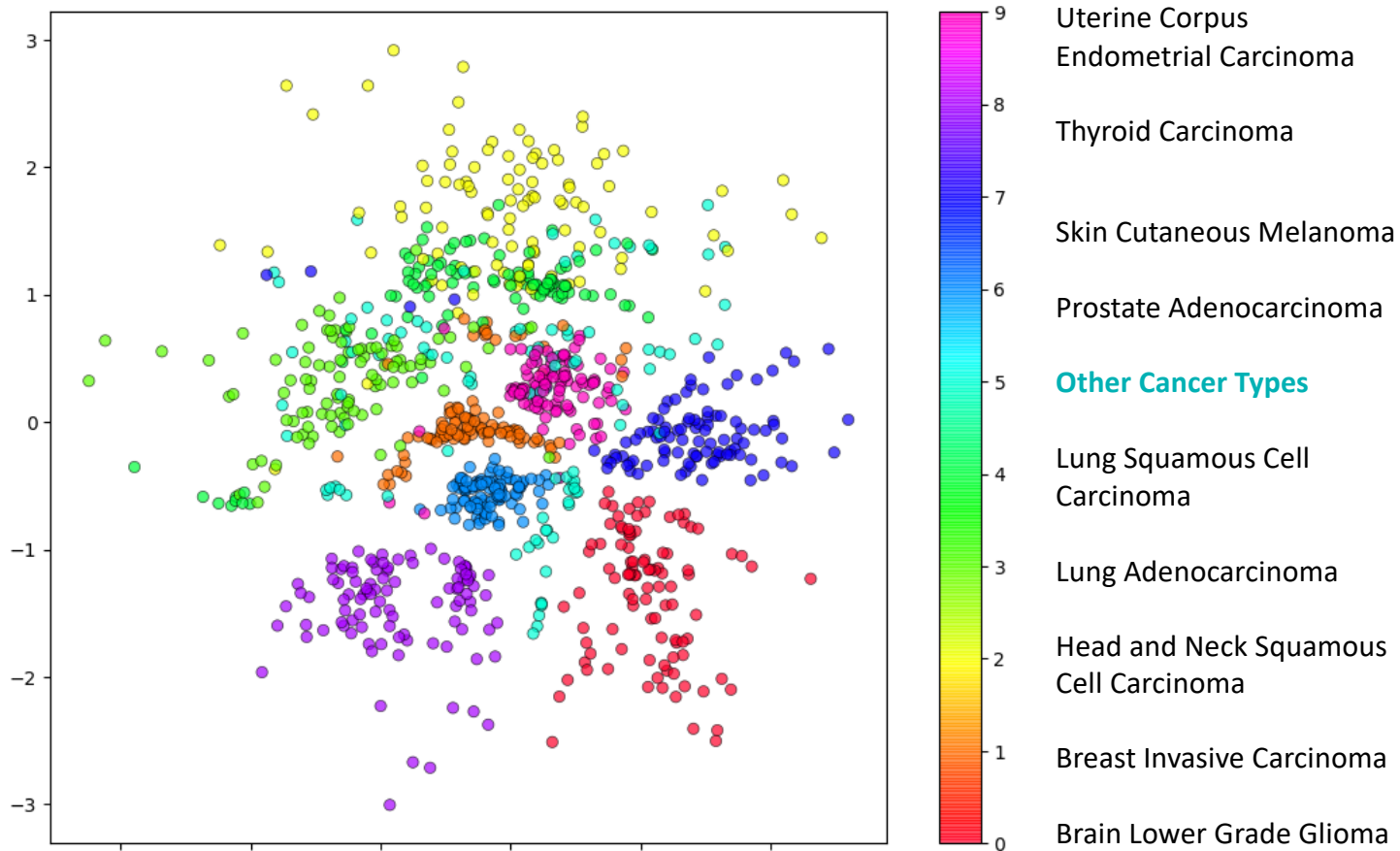


<https://github.com/ECP-CANDLE/Benchmarks/tree/frameworks/Pilot1/TC1>



<https://github.com/ECP-CANDLE/Benchmarks/tree/frameworks/Pilot1/TC1>

VAE Latent Representation of GDC Expression



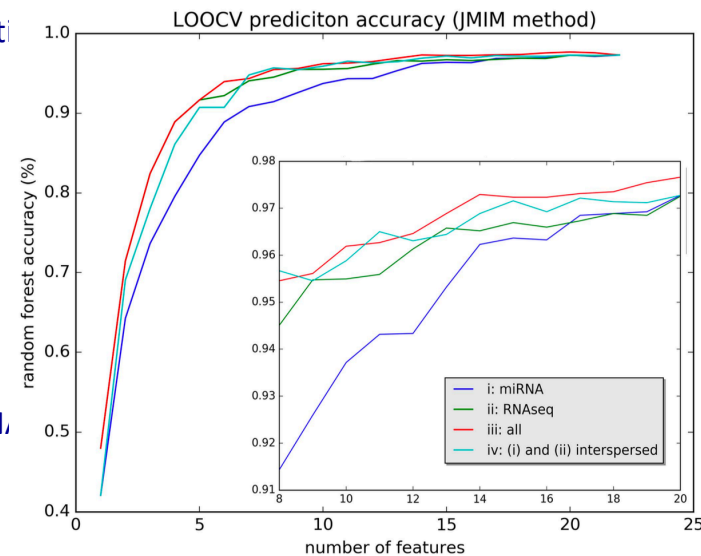
<https://github.com/ECP-CANDLE/Benchmarks/tree/frameworks/Pilot1/P1B1>

How did we know it might work?

- Build autoencoders first with the features you are going to work with
- If you get reasonable accuracy then the model can learn a representation and that is a good sign
- Class balance seems to matter
- Number of training examples matters > 1000 is good > 10,000 better, > 100,000 much better
- Hyper parameter search is also important once you get something that basically works

Generate Compact Molecular Signatures

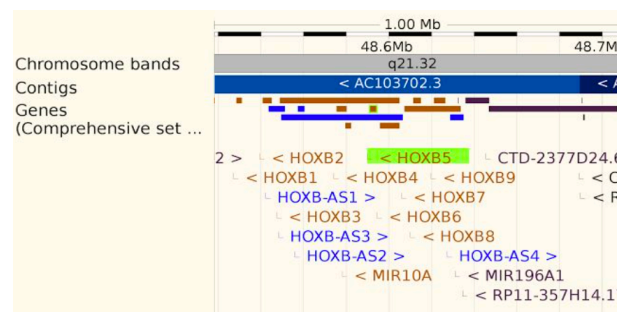
- **For each agent or class of agents we will apply feature selection methods to the models to generate where possible a compact molecular signature that retains prediction performance**
 - Typical reduced signatures include $O(10)$ - $O(100)$ features from $\gg 50,000$ starting features
 - Features may be genes, SNPs, μ RNA etc.
- **Developed and applied multiple feature selection methods**
 - Selection criterion: Chi2, Anova, mutual info, ensemble ML, deep neural networks
 - Algorithms: ranking, intersection, iterative maximization
 - Supervised recursive binning
- **Extracted compact features**
 - Features from cancer type prediction
 - 50 features: 0.981 accuracy
 - 20 features: 0.976 accuracy
 - 14 features: 0.973 accuracy
 - RNAseq is more informative than miRN.



Analyze Molecular Signatures to Provide Insight to Potential Mechanisms

- **Started mapping gene features to pathways**
 - Enrichment analysis will be applied to the signatures to identify associated pathways
 - Pathways will be identified that associate with both sensitive and resistant response phenotypes
- **Identified co-located or known interacting pairs of gene and microRNA signatures**

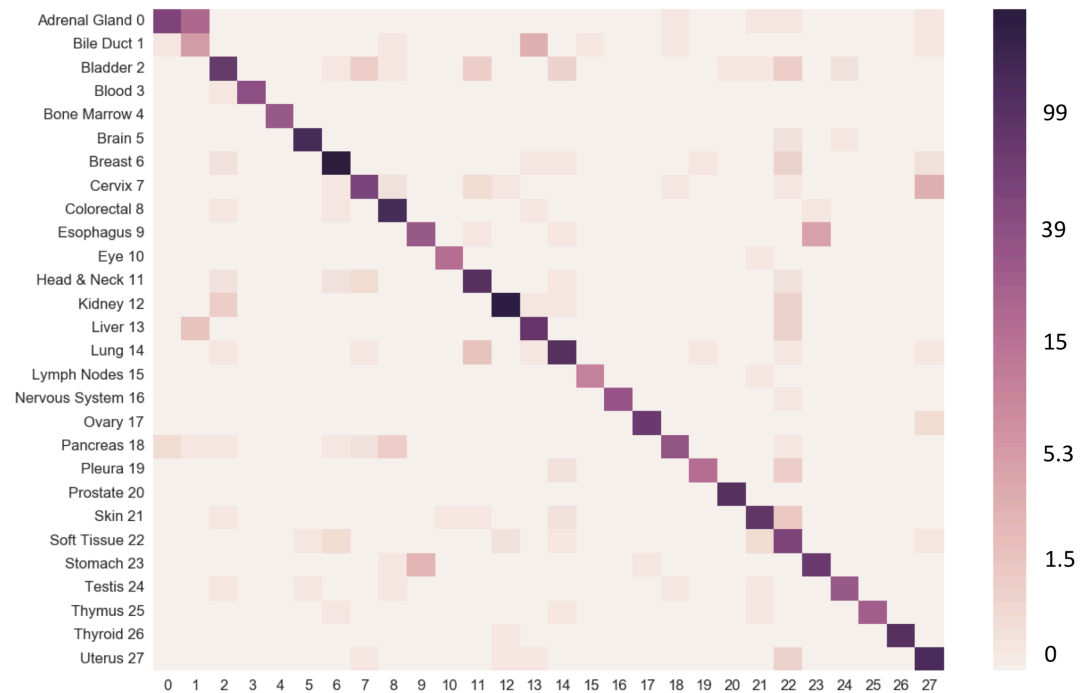
- Top miRNA feature hsa.mir.10a is co-located with ENSG00000120075.5
- It has also been experimentally verified that this miRNA downregulates the corresponding HOX genes



Rank	MIR	RNA	MIR&RNA
1	hsa.mir.10a	ENSG00000119888.9	ENSG00000119888.9
2	hsa.mir.205	ENSG00000170370.11	ENSG00000170370.11
3	hsa.mir.181a.2	ENSG00000157551.16	ENSG00000157551.16
4	hsa.mir.135a.1	ENSG00000124664.9	ENSG00000124664.9
5	hsa.mir.203a	ENSG00000102554.12	ENSG00000102554.12
6	hsa.mir.196b	ENSG0000009765.13	hsa.mir.10a
7	hsa.mir.194.1	ENSG00000275410.3	ENSG0000009765.13
8	hsa.mir.9.3	ENSG00000274173.1	ENSG00000275410.3
9	hsa.mir.196a.2	ENSG00000120075.5	ENSG00000274173.1
10	hsa.mir.429	ENSG00000124466.8	hsa.mir.205
11	hsa.mir.375	ENSG00000204385.9	ENSG00000204385.9
12	hsa.mir.584	ENSG00000103449.10	ENSG00000104447.10
13	hsa.mir.135b	ENSG00000104447.10	ENSG00000103449.10
14	hsa.mir.10b	ENSG00000255794.5	ENSG00000078399.14
15	hsa.let.7i	ENSG00000189334.7	ENSG00000189334.7
16	hsa.mir.125b.2	ENSG00000165215.6	ENSG00000165215.6
17	hsa.mir.30a	ENSG00000137203.9	ENSG00000137203.9
18	hsa.mir.200c	ENSG00000078399.14	ENSG00000255794.5
19	hsa.mir.203b	ENSG00000103942.11	ENSG00000103942.11
20	hsa.mir.944	ENSG00000046653.13	ENSG00000046653.13
21	hsa.mir.1301	ENSG00000151322.17	ENSG00000151322.17
22	hsa.mir.138.1	ENSG00000123892.10	ENSG00000123892.10

Close examination of prediction error

- Confusion matrix
- Local feature importance
- Force plots



RNAseq Bias Removal

Alex Partin

*Cancer Type Classification
with SNPs*

GDC: 10K samples with 10M mutations

VEP Impact

- MODIFIER 2,304,599
- MODERATE 1,666,904
- LOW 714,906
- HIGH 292,356

SIFT Impact

- deleterious 851,323
- tolerated 677,244
- deleterious_low_confidence 137,952
- tolerated_low_confidence 102,495

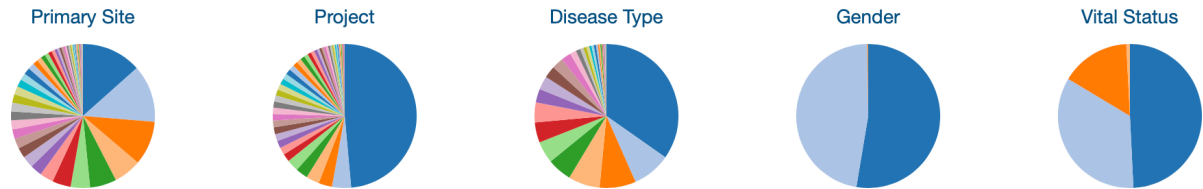
Polyphen Impact

- benign 806,244
- probably_damaging 675,545
- possibly_damaging 464,623
- unknown 101,253

Consequence Type

- missense_variant 1,648,416
- non_coding_transcript_exon_variant 1,119,774
- downstream_gene_variant 1,069,273

Cases (37,075) Genes (22,872) Mutations (3,142,246) OncoGrid



Showing 1 - 20 of 37,075 cases

Download Biospecimen Clinical JSON TSV Save/Edit Case Set

Case ID	Project	Primary Site	Gender	Files	Available Files per Data Category							# Mutations	# Genes	Slides
					Seq	Exp	SNV	CNV	Meth	Clinical	Bio			
TCGA-A5-A0G2	TCGA-UCEC	Corpus uteri	Female	58	4	5	16	5	1	10	17	42,051	14,357	h (3)
TCGA-EO-A22U	TCGA-UCEC	Corpus uteri	Female	57	4	5	16	5	1	10	16	26,998	12,629	h (2)
TCGA-FI-A2D5	TCGA-UCEC	Corpus uteri	Female	58	4	5	16	5	1	11	16	26,139	12,482	h (2)
TCGA-AX-A2HC	TCGA-UCEC	Corpus uteri	Female	65	6	10	16	5	2	10	16	24,853	12,205	h (2)
TCGA-EO-A22R	TCGA-UCEC	Corpus uteri	Female	59	4	5	16	5	2	10	17	24,276	11,920	h (3)
TCGA-B5-A3FC	TCGA-UCEC	Corpus uteri	Female	57	4	5	16	5	1	10	16	24,584	11,902	h (2)
TCGA-IB-7651	TCGA-PAAD	Pancreas	Female	56	4	5	16	5	1	8	17	23,084	11,453	h (3)
TCGA-AP-A1DV	TCGA-UCEC	Corpus uteri	Female	55	4	5	16	3	1	10	16	22,664	11,431	h (2)
TCGA-E6-A1LX	TCGA-UCEC	Corpus uteri	Female	57	4	5	16	5	1	10	16	23,542	11,397	h (2)
TCGA-AP-A0LM	TCGA-UCEC	Corpus uteri	Female	58	4	5	16	5	1	10	17	22,507	11,301	h (3)
TCGA-2W-A8YY	TCGA-CESC	Cervix uteri	Female	56	4	5	16	5	1	9	16	21,749	11,177	h (2)
TCGA-AX-A1CE	TCGA-UCEC	Corpus uteri	Female	58	4	5	16	5	1	10	17	21,720	11,153	h (3)
TCGA-AP-A1DK	TCGA-UCEC	Corpus uteri	Female	57	4	5	16	5	1	10	16	20,472	11,059	h (2)
TCGA-A5-A1OF	TCGA-UCEC	Corpus uteri	Female	57	4	5	16	5	1	10	16	20,080	10,990	h (2)

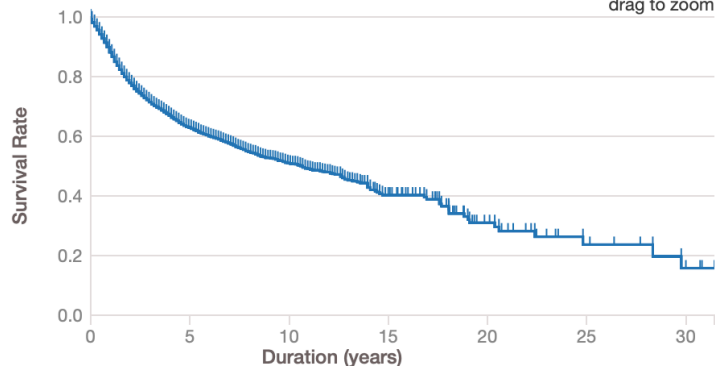
Somatic Mutations

Overall Survival Plot



17,424 Cases with Survival Data

drag to zoom



Showing 1 - 10 of 3,142,246 somatic mutations



<input type="checkbox"/> DNA Change	Type	Consequences	# Affected Cases in Cohort	# Affected Cases Across the GDC	Impact	Survival
<input type="checkbox"/> chr7:g.140753336A>T	Substitution	Missense BRAF V600E	565 / 10,202 5.54%	565 / 10,202	MO DH PR	
<input type="checkbox"/> chr2:g.208248388C>T	Substitution	Missense IDH1 R132H	388 / 10,202 3.80%	388 / 10,202	MO DL PO	
<input type="checkbox"/> chr3:g.179218303G>A	Substitution	Missense PIK3CA E545K	258 / 10,202 2.53%	258 / 10,202	MO DH PR	
<input type="checkbox"/> chr3:g.179234297A>G	Substitution	Missense PIK3CA H1047R	234 / 10,202 2.29%	234 / 10,202	MO TO PO	
<input type="checkbox"/> chr12:g.25245350C>T	Substitution	Missense KRAS G12D	208 / 10,202 2.04%	208 / 10,202	MO DH BE	
<input type="checkbox"/> chr12:g.25245350C>A	Substitution	Missense KRAS G12V	176 / 10,202 1.73%	176 / 10,202	MO DH PO	
<input type="checkbox"/> chr3:g.179218294G>A	Substitution	Missense PIK3CA E542K	167 / 10,202 1.64%	167 / 10,202	MO DH PR	
<input type="checkbox"/> chr17:g.7675088C>T	Substitution	Missense TP53 R175H	156 / 10,202 1.53%	156 / 10,202	MO TO BE	
<input type="checkbox"/> chr17:g.7673803G>A	Substitution	Missense TP53 R273C	125 / 10,202 1.23%	125 / 10,202	MO DH PR	

How to deal with $n \gg p$?

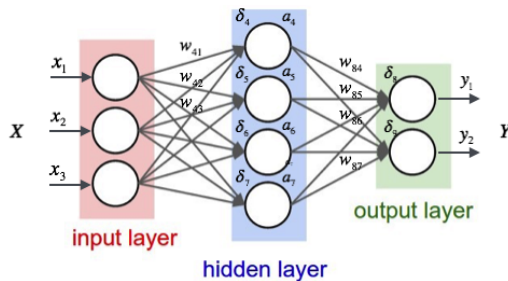
- Dropout
- Regularization
- Locally connected networks

Diet Networks: Thin Parameters for Fat Genomics

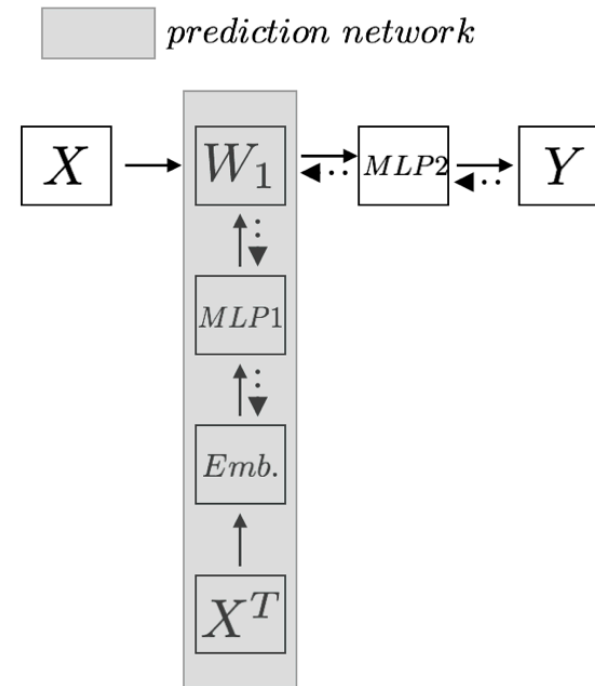
Adriana Romero, Pierre Luc Carrier, Akram Erraqabi, Tristan Sylvain, Alex Auvolat, Etienne Dejoie, Marc-André Legault, Marie-Pierre Dubé, Julie G. Hussin, Yoshua Bengio

Diet Network

- Suppose we have
 - 1000 samples
 - 1,000,000 features
 - 100 neurons in the hidden layer

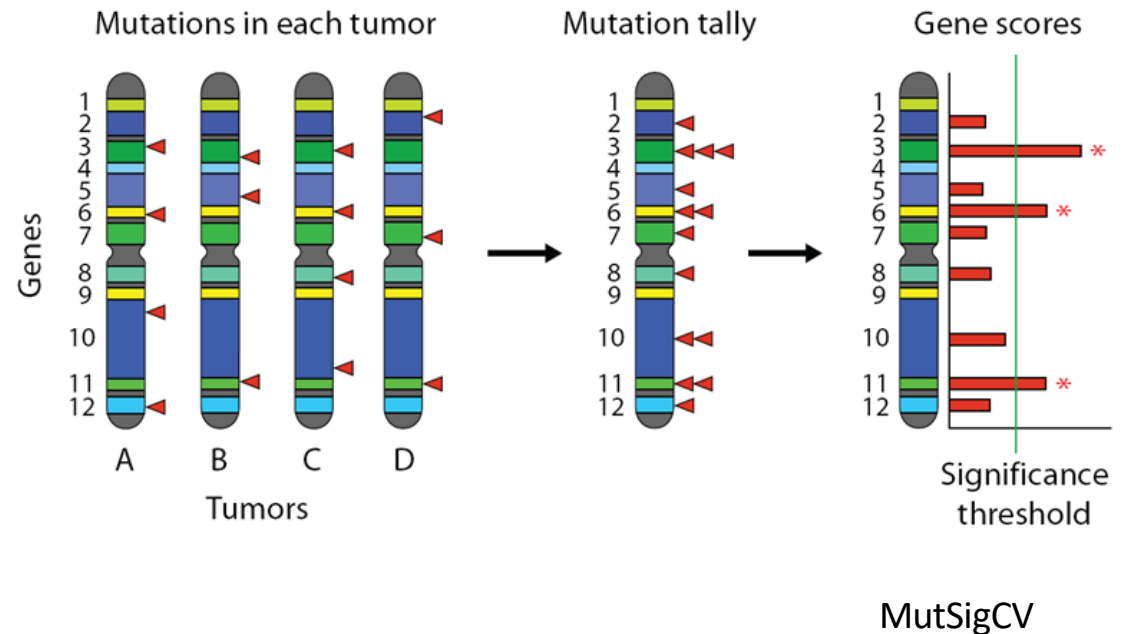


- Parameters in first layer = 100M

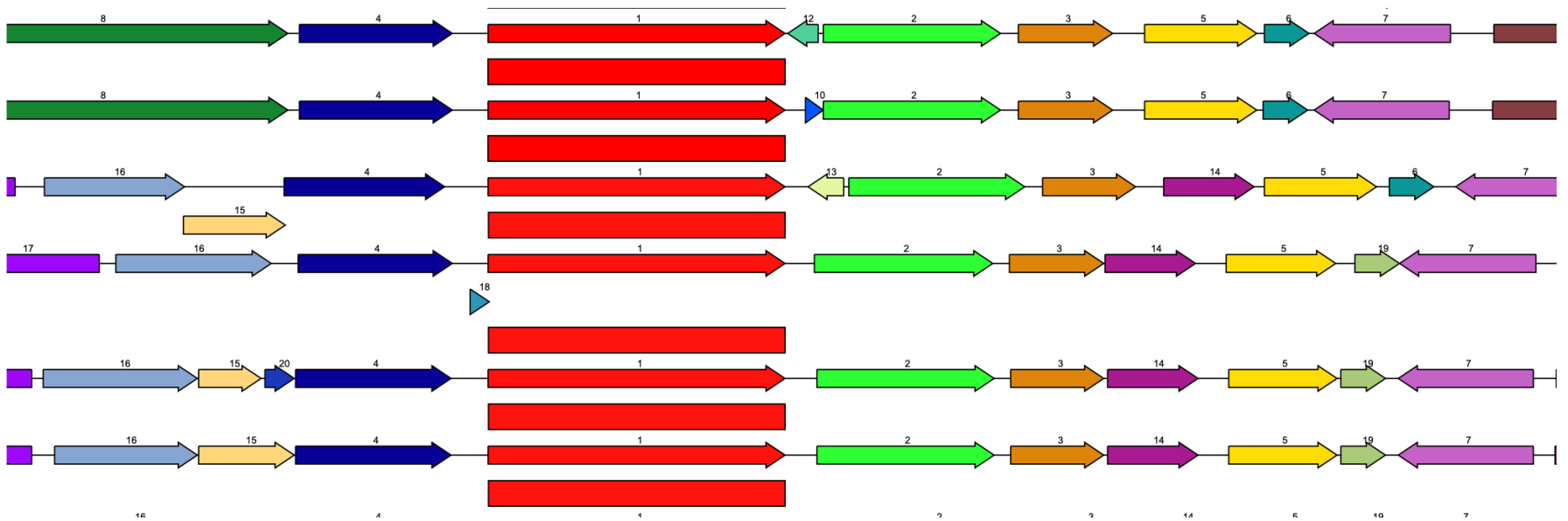


How do we represent the sparse mutations?

- Gene level / pathway level
- Weighting by impact
- Filtering by significance
- Convert to images
 - Variant calling
 - Annotation



Deep annotation with compare region images



Identification of genomic islands and operons

Table 3. Quality of tools predictions: % predictions made with GI features | % predictions missed with GI features, over the testing genomes dataset.

Predictor \ Target	ShutterIsland	IslandViewer	AlienHunter	IslandPick	SIGI	Average
ShutterIsland	N/A	91% 64%	87% 47%	89% 31%	87% 36%	89% 45%
IslandViewer	94% 67%	N/A	89% 45%	80% n/a	87% n/a	88% 56%
AlienHunter	74% 70%	66% 60%	N/A	73% 21%	71% 42%	71% 48%
IslandPick	69% 76%	34% 86%	49% 53%	N/A	54% 44%	52% 65%
SIGI	67% 75%	45% 77%	48% 51%	50% 35%	N/A	53% 60%
Dimob	n/a 66%	n/a 28%	n/a 43%	n/a 25%	n/a 23%	n/a 37%
Phispy	n/a 68%	n/a 70%	n/a 50%	n/a 33%	n/a 39%	n/a 52%
PhageFinder	n/a 68%	n/a 70%	n/a 50%	n/a 34%	n/a 39%	n/a 52%
Islander	n/a 75%	n/a 71%	n/a 51%	n/a 33%	n/a 39%	n/a 54%
Phaster	n/a 75%	n/a 71%	n/a 51%	n/a 33%	n/a 39%	n/a 54%

WORD2VEC

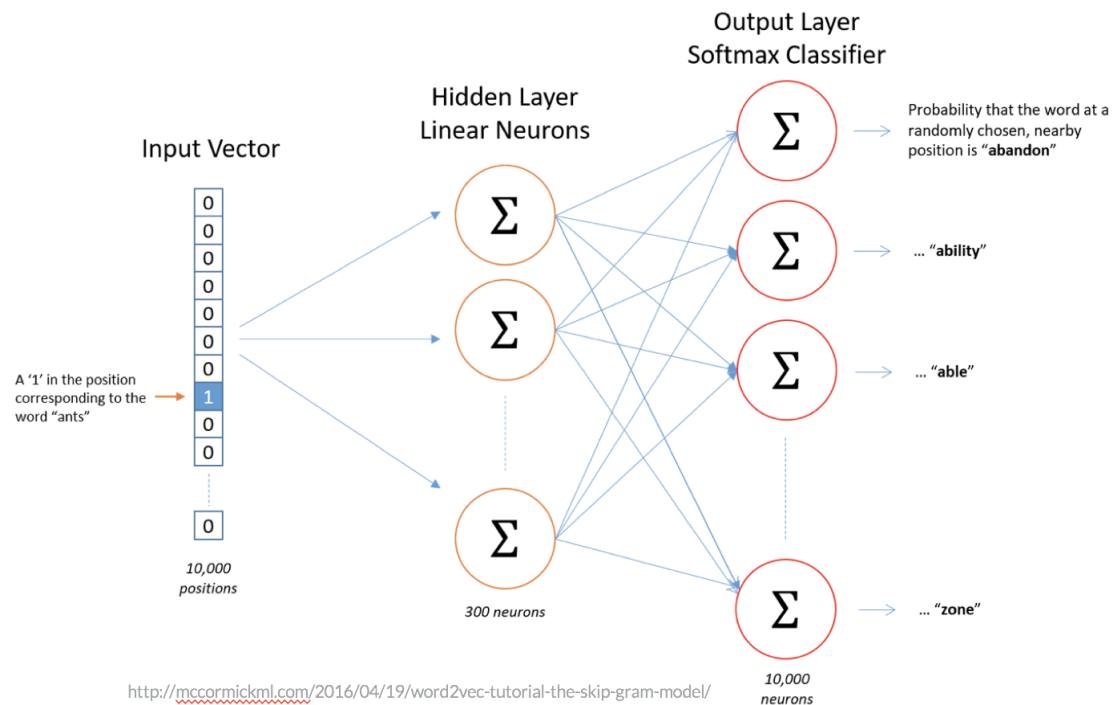
WINDOW

THE QUICK BROWN FOX JUMPS OVER THE LAZY DOG



CLASSIFIERS

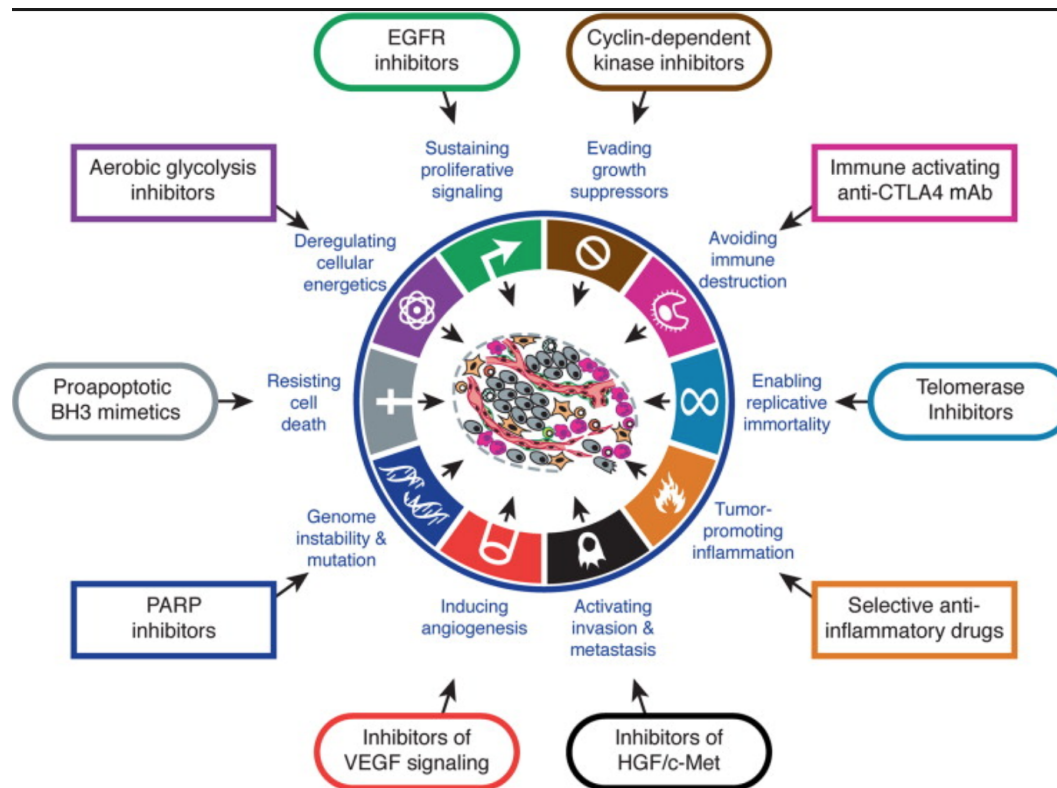




“Skip-Gram”
 With one-hot encoded centre word, we can predict context words.

Hidden layer creates embeddings

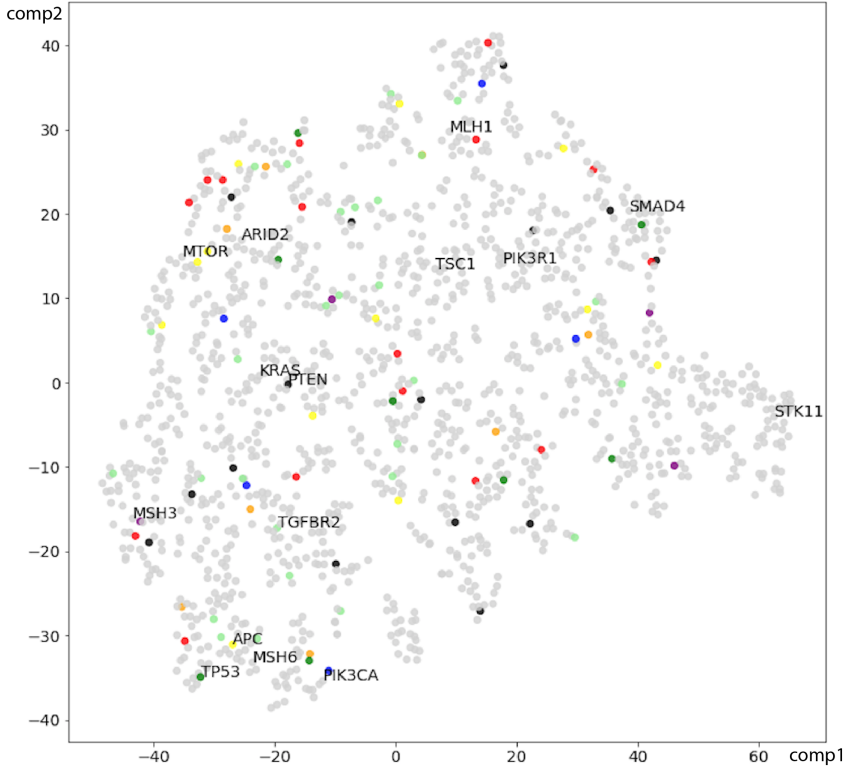
Gene sets from MSigDB



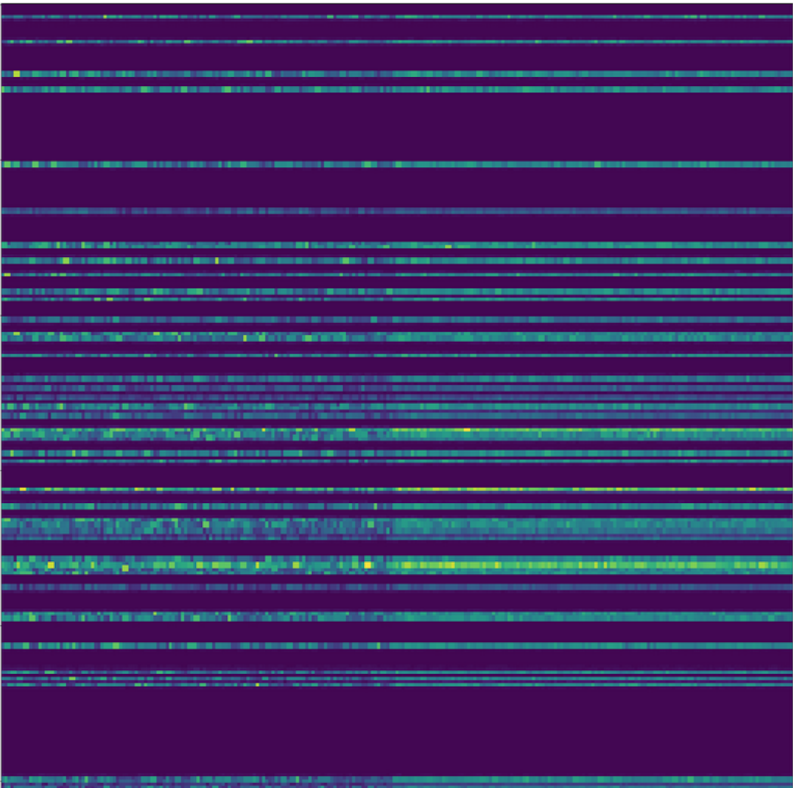
22,596 gene sets

- H** **hallmark gene sets** are coherently expressed signatures derived by aggregating many MSigDB gene sets to represent well-defined biological states or processes.
- C1** **positional gene sets** for each human chromosome and cytogenetic band.
- C2** **curated gene sets** from online pathway databases, publications in PubMed, and knowledge of domain experts.
- C3** **motif gene sets** based on conserved cis-regulatory motifs from a comparative analysis of the human, mouse, rat, and dog genomes.
- C4** **computational gene sets** defined by mining large collections of cancer-oriented microarray data.
- C5** **GO gene sets** consist of genes annotated by the same GO terms.
- C6** **oncogenic gene sets** defined directly from microarray gene expression data from cancer gene perturbations.
- C7** **immunologic gene sets** defined directly from microarray gene expression data from immunologic studies.

Gene2vec



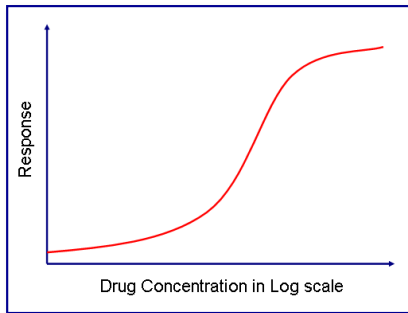
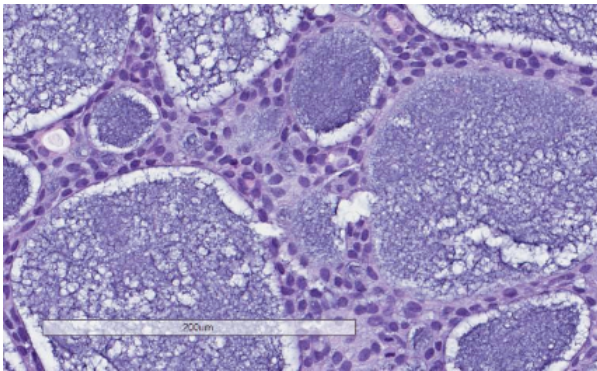
Sample2image



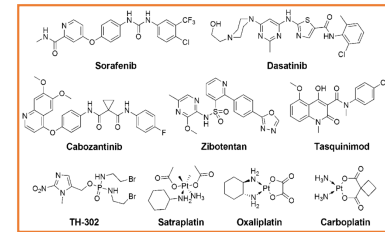
Alena Harley, The Mystery of the Origin

Drug Response Prediction

Modeling Drug Response



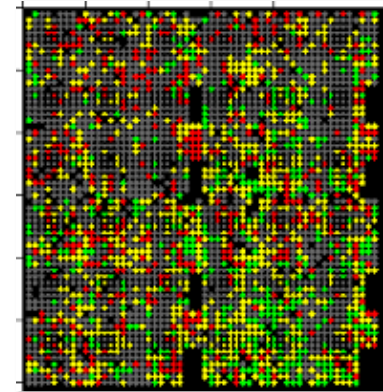
Drug (s)
descriptors
fingerprints
structures
SMILES
dose



$$\mathcal{R} = f(\mathcal{T}, \mathcal{D}_1, \mathcal{D}_2)$$

↑
IC50
GI50
% growth
Z-score
AUC
Response

↑
gene expression levels
SNPs
protein abundance
microRNA
methylation
Tumor



Cell Line Features

- NCI-60: 60 cell lines
- Molecular Assays: 20
 - Gene expression array
 - RNA-seq
 - Mutations
 - Protein abundance
 - microRNA

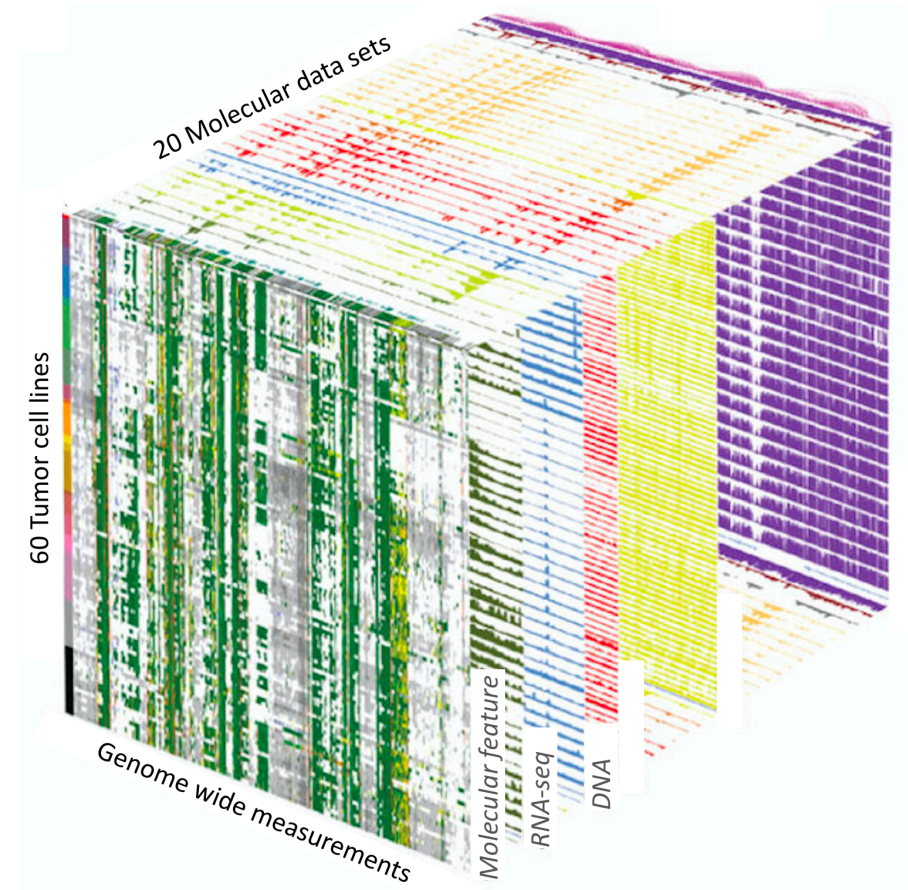
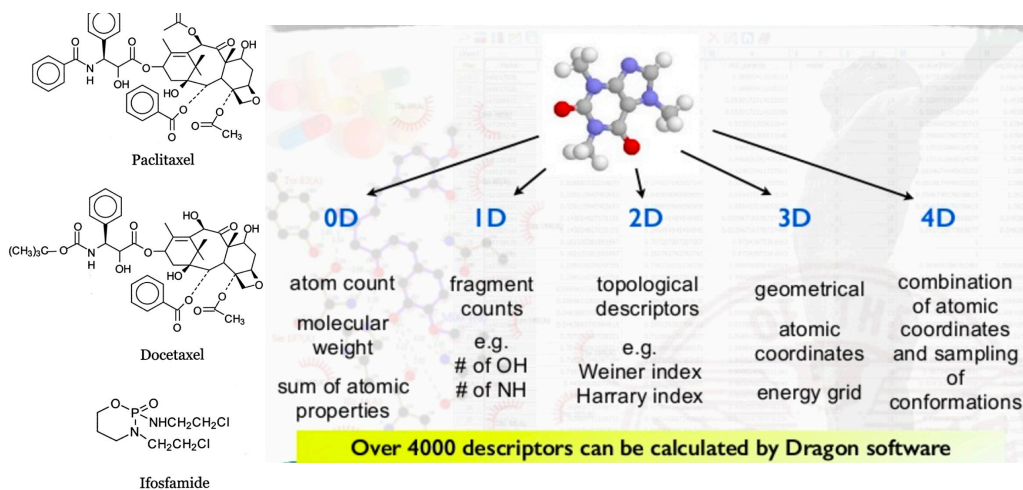
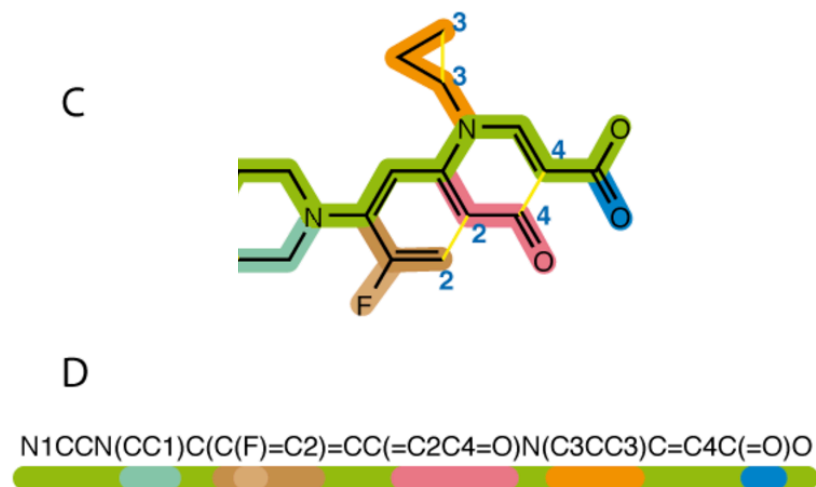
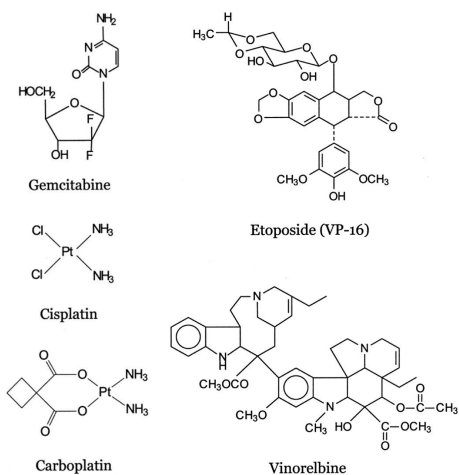


Figure adapted from Kundaje et al. Nature 2015

Drug Features

- SMILES strings
- 2D or 3D structures
- Graph convolutions
- Descriptors
- Fingerprints



Deep Learning Model for Drug Pair Response

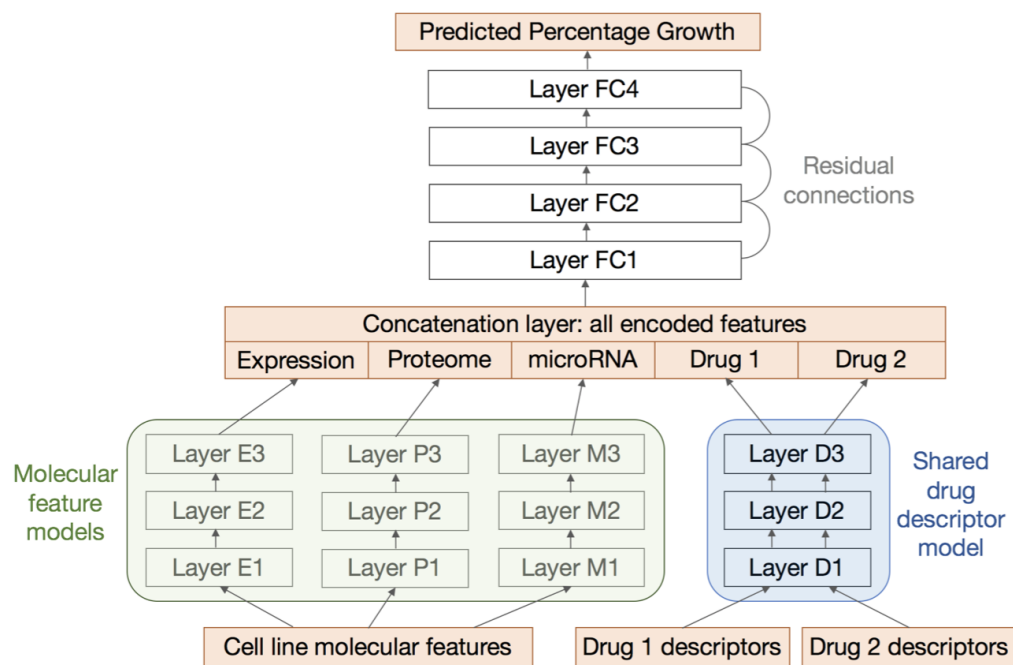
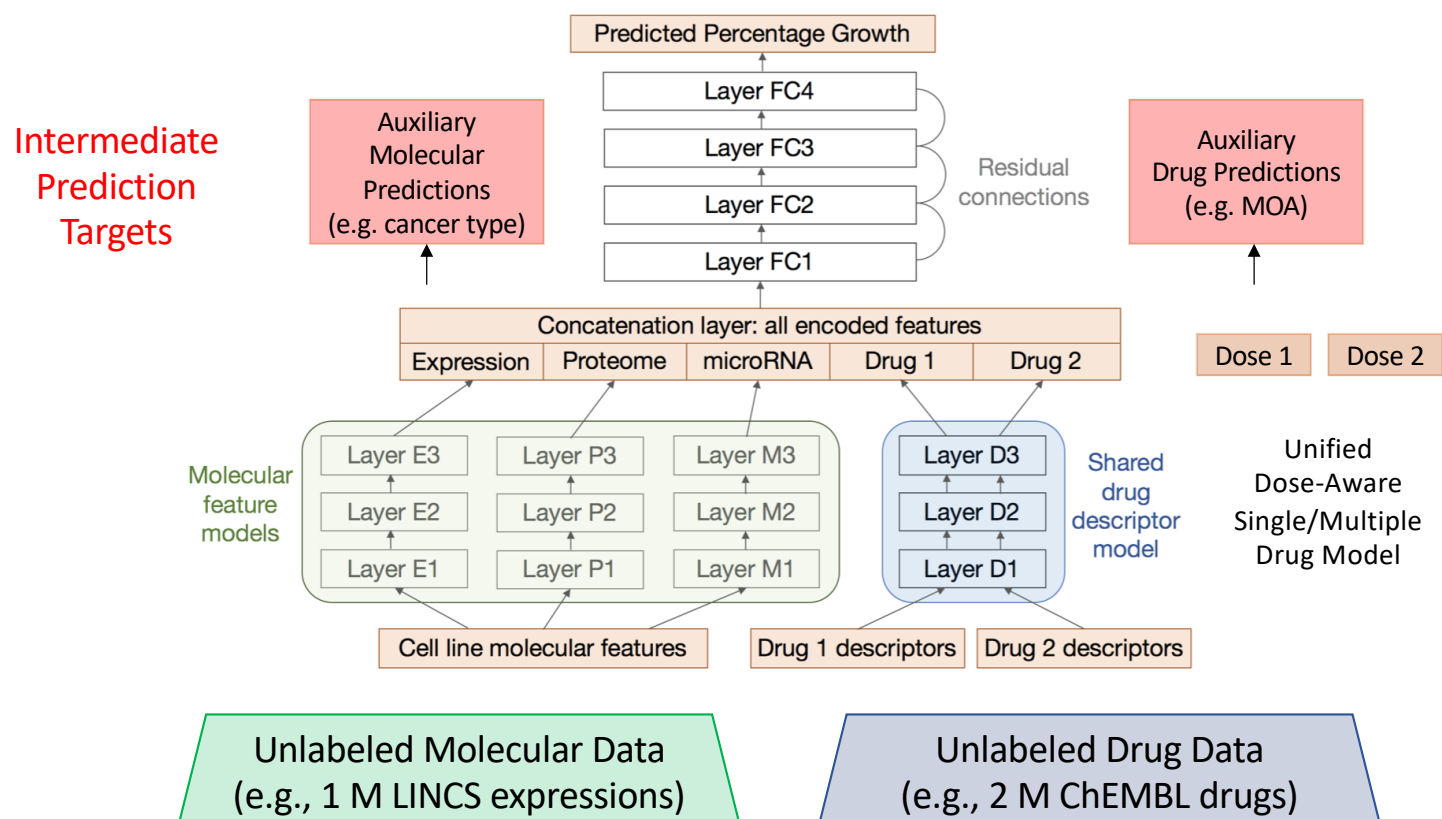
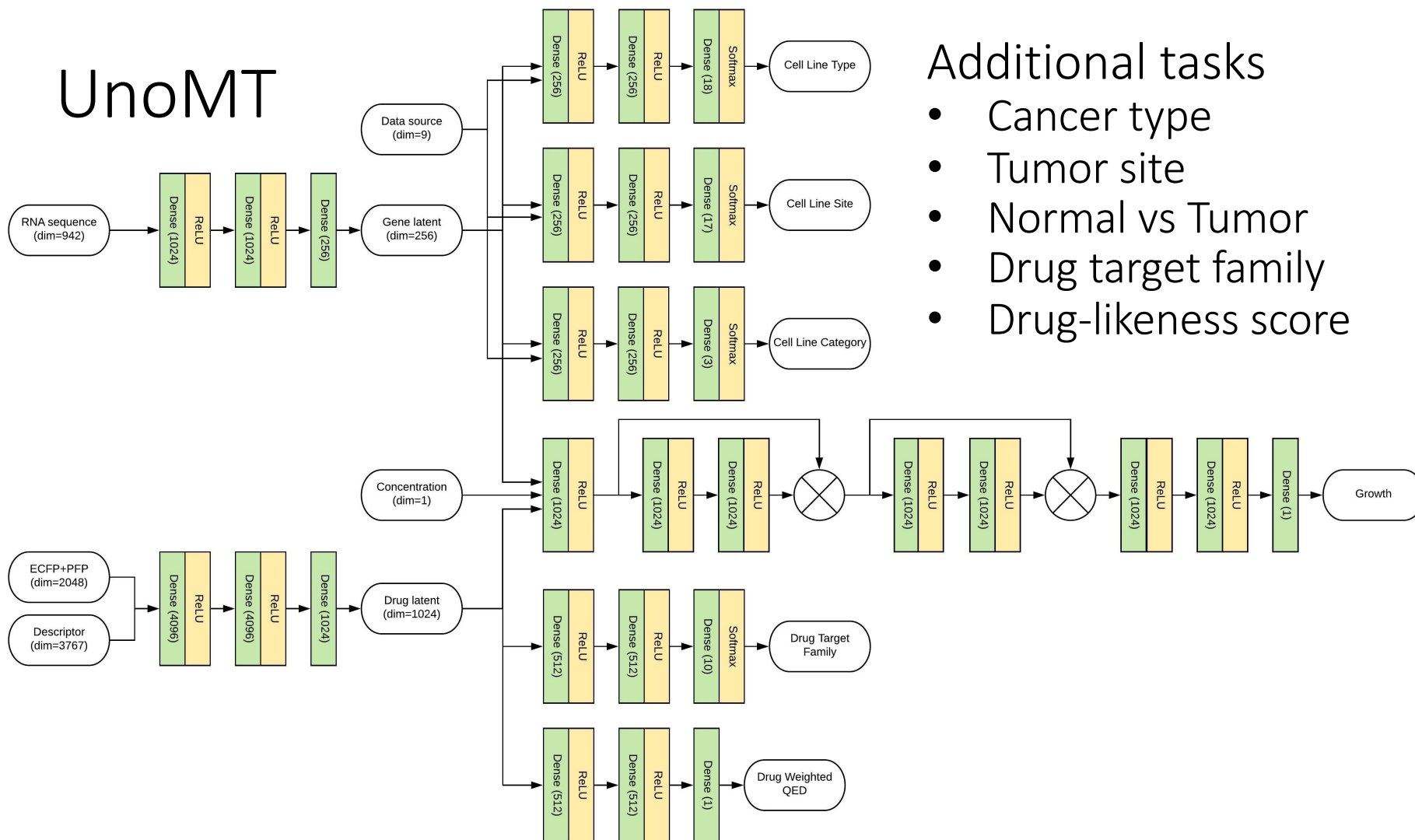


Fig. 2. **Neural network architecture.** The orange square boxes, from bottom to top, represent input features, encoded features, and output growth values. Feature models are denoted by round shaded boxes: green for molecular features and blue for drug features. There are multiple types of molecular features that are fed into submodels for gene expression, proteome, and microRNA. The descriptors for the two drugs share the same descriptor model. All encoded features are then concatenated to form input for the top fully connected layers. Most connecting layers are linked by optional residual skip connections if their dimensions match.

Uno: Predicting Single/Paired Drug Response

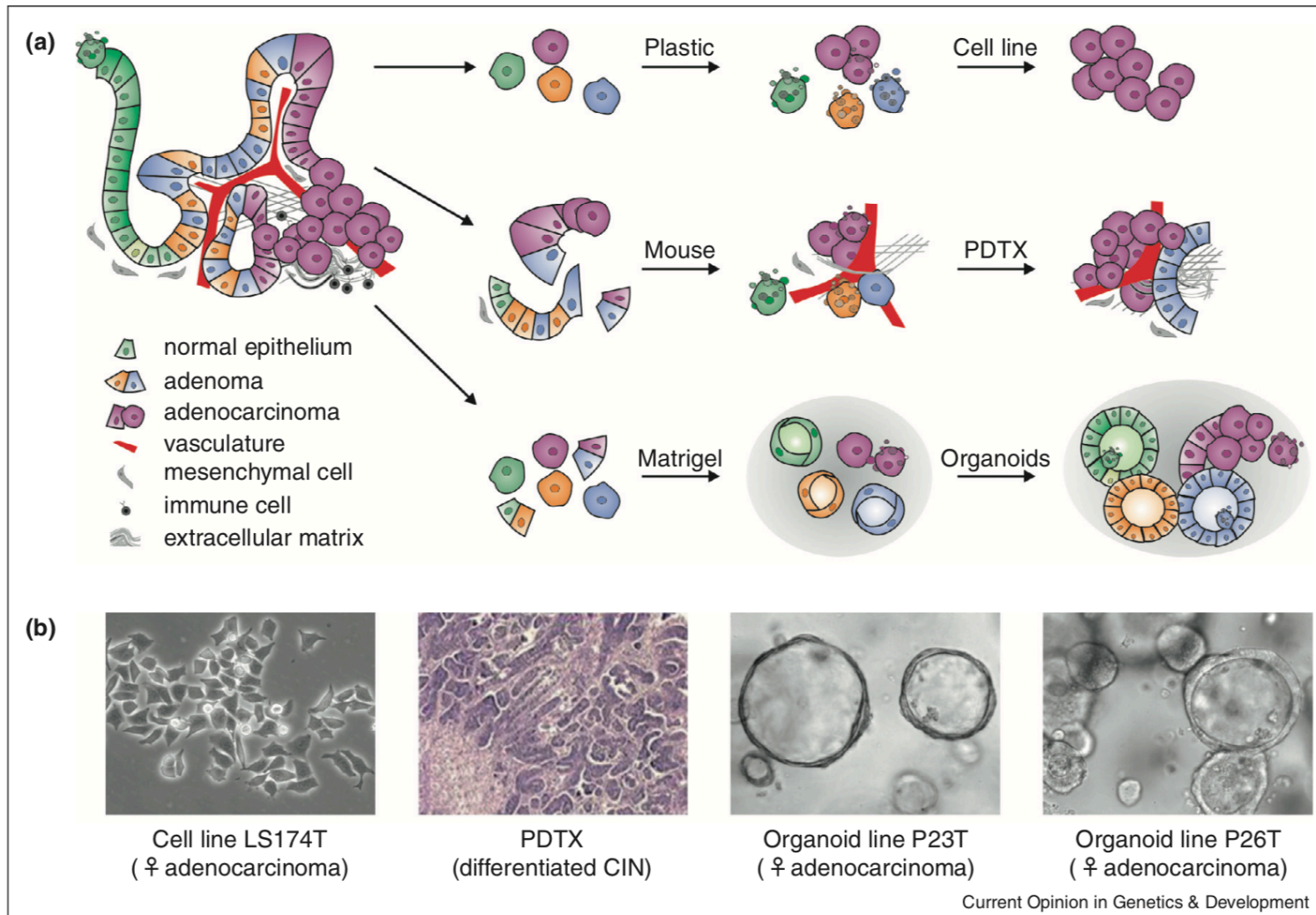


UnoMT



Additional tasks

- Cancer type
- Tumor site
- Normal vs Tumor
- Drug target family
- Drug-likeness score



Organoid cultures for the analysis of cancer phenotypes, Sachs and Clevers, 2014

Pilot 1

Data in place for model training and testing

Table 1. Integrating cell line, PDX, and real tumor samples across multiple studies

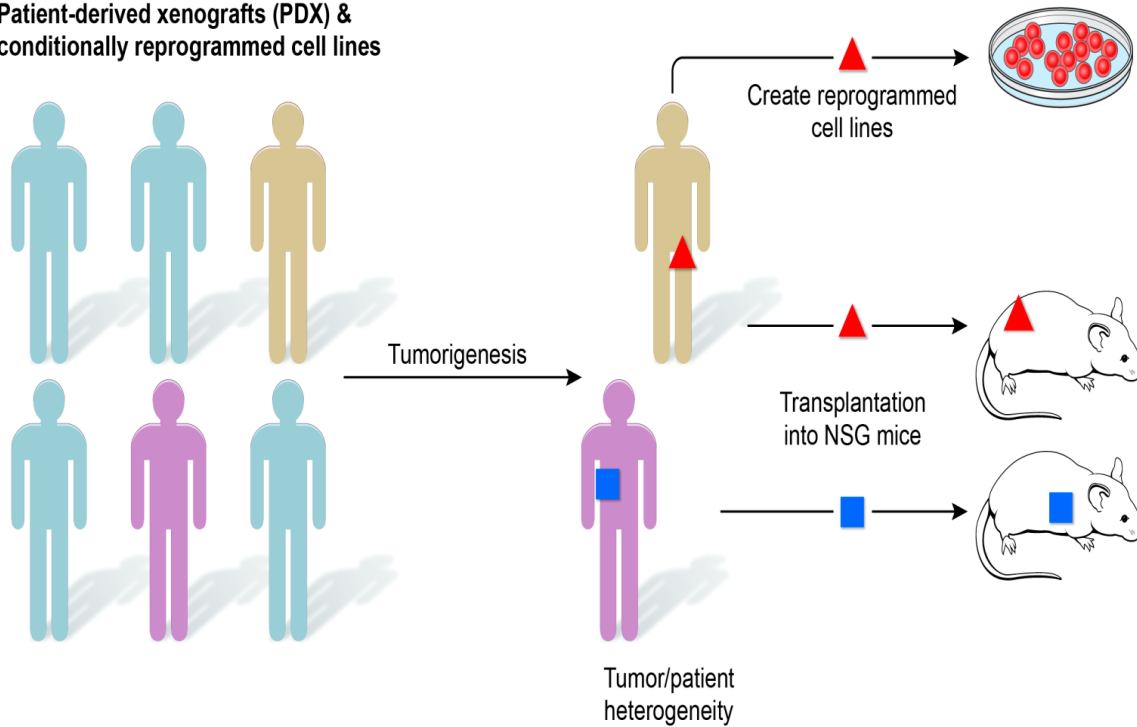
Dose Independent	Data Source	# Tumor Samples	# Drugs	# Dose Response Samples	Treatment Type
11,671	NCI-ALMANAC	60	104	3,686,475	Drug pair
395,264	CCLE	504	24	93,251	Single drug
6,456	CTRPv2	887	544	6,171,005	Single drug
225,481	gCSI	409	16	58,094	Single drug
3,780,150 (60,000)	GDSC	1,075	249	1,894,212	Single drug
	NCI	60	52,671	18,862,308	Single drug
	GDC	11,081	N/A	N/A	N/A
	NCI-PDM	1,198	12	518*	Single and paired drugs

* PDM drug response were measured differently from cell line dose response data.

Pilot 1

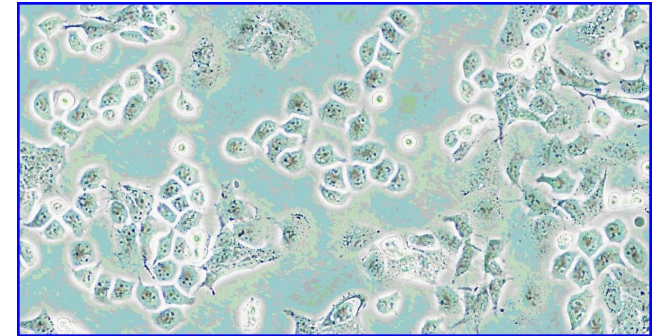
Patient Derived Xenograft Models

Patient-derived xenografts (PDX) & conditionally reprogrammed cell lines



Nature Rev. Clin. Oncol. 11: 649-662, 2014.

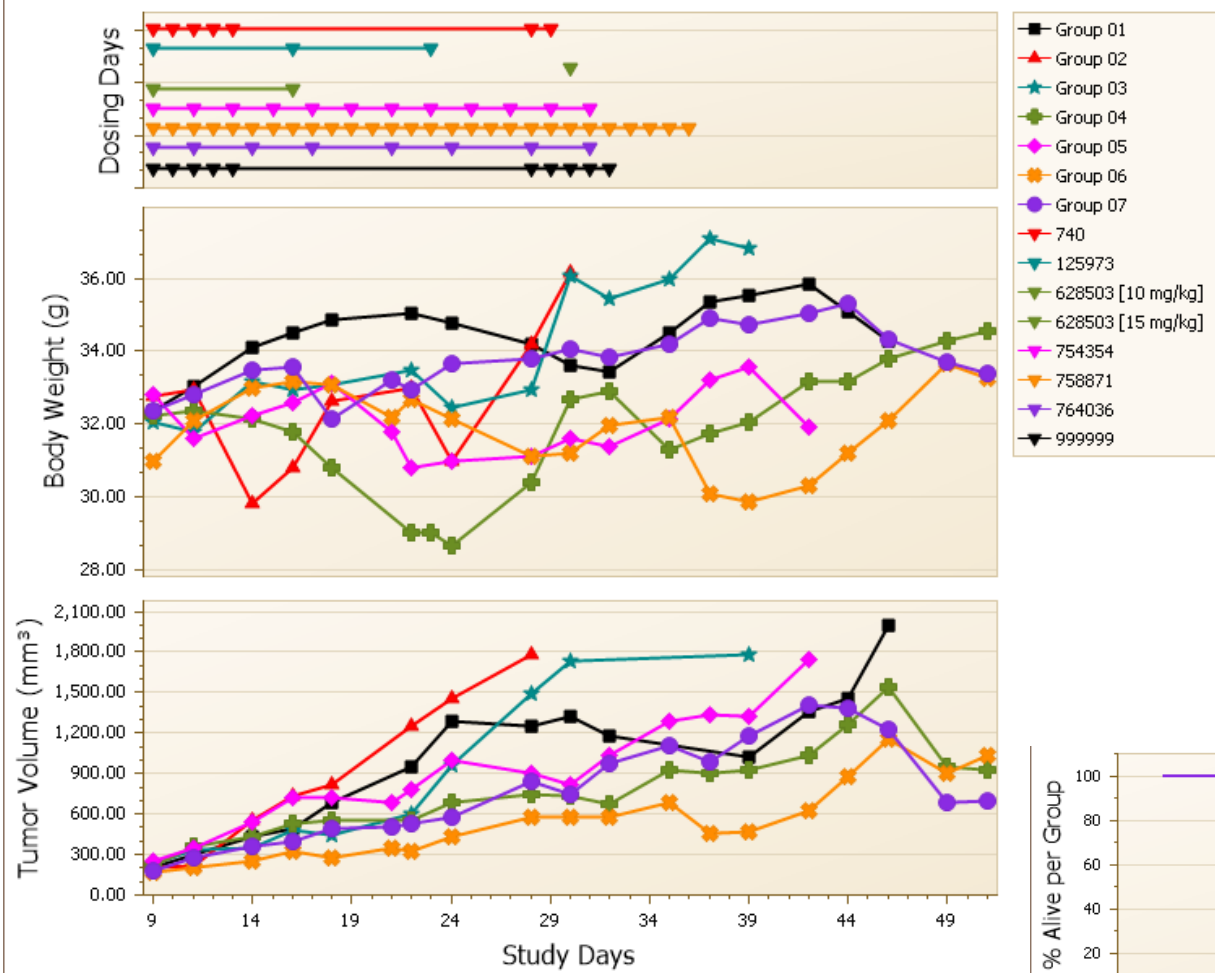
Cancer Cell Lines



CL and PD Xenografts



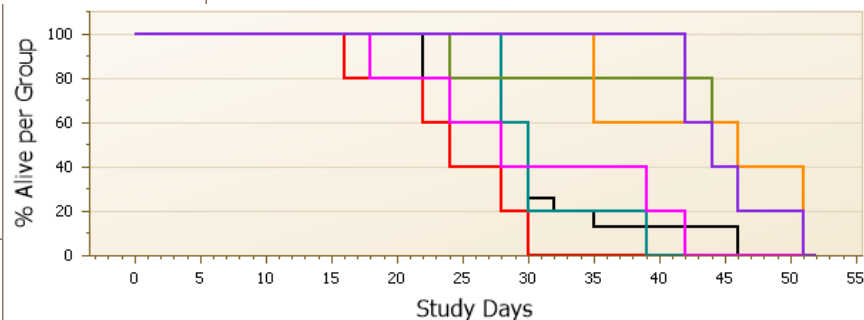
ZFZJ2-1-P-8B



ZFZJ2-1, 146476-266-R, Urothelial/bladder Ca

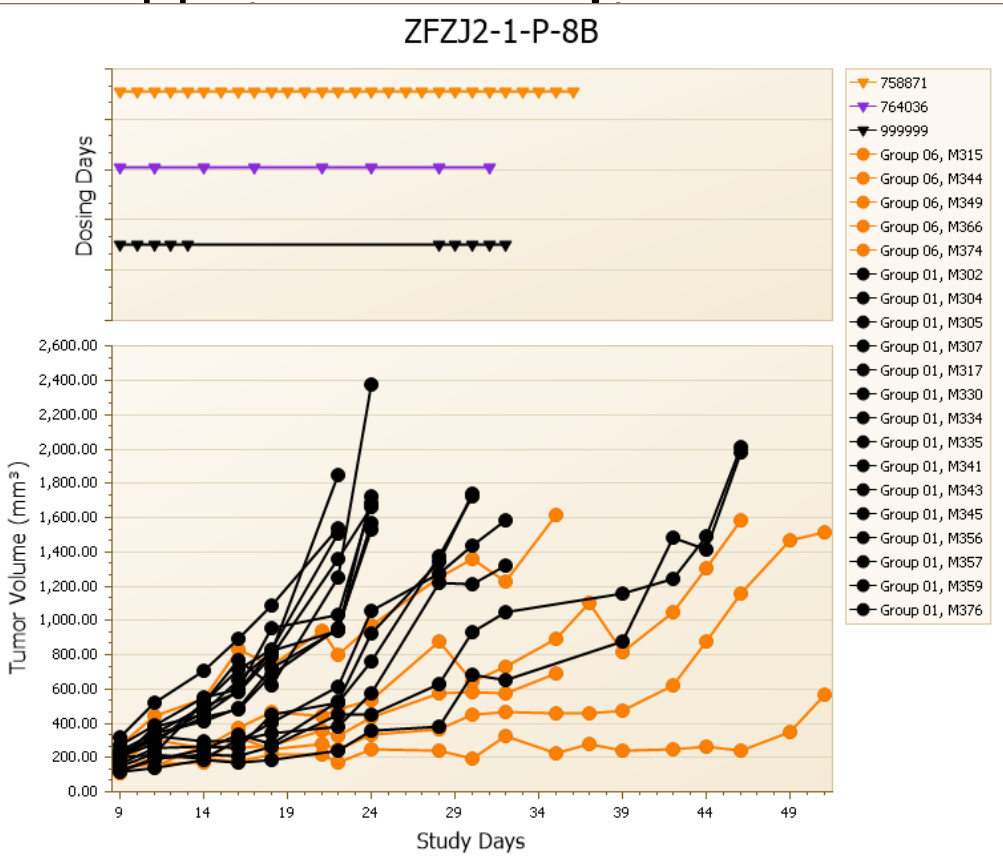
AZD8055 ~2.2x delay. No regression

Paclitaxel	125973
AZD8055	758871
Dinaciclib	764036
Docetaxel	628503
GSK-461364	754354
Methotrexate	740

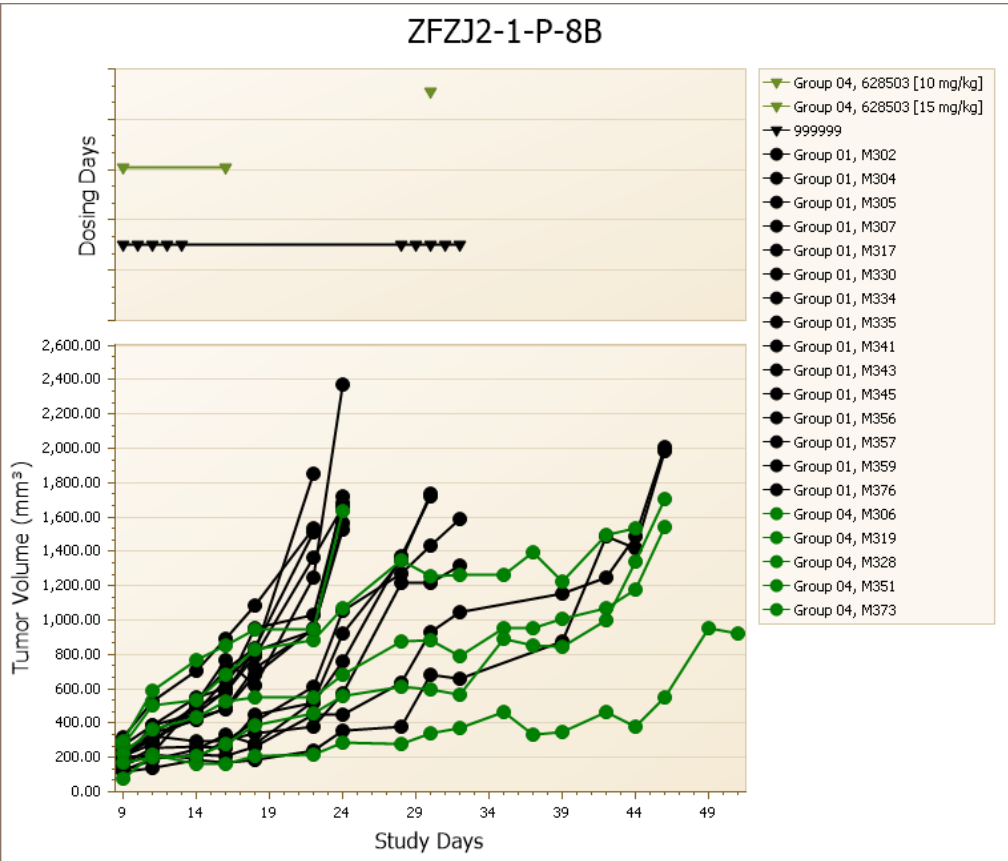


ZFZJ2-1, 146476-266-R, Urothelial/bladder ca

ZFZJ2-1-P-8B



ZFZJ2-1-P-8B

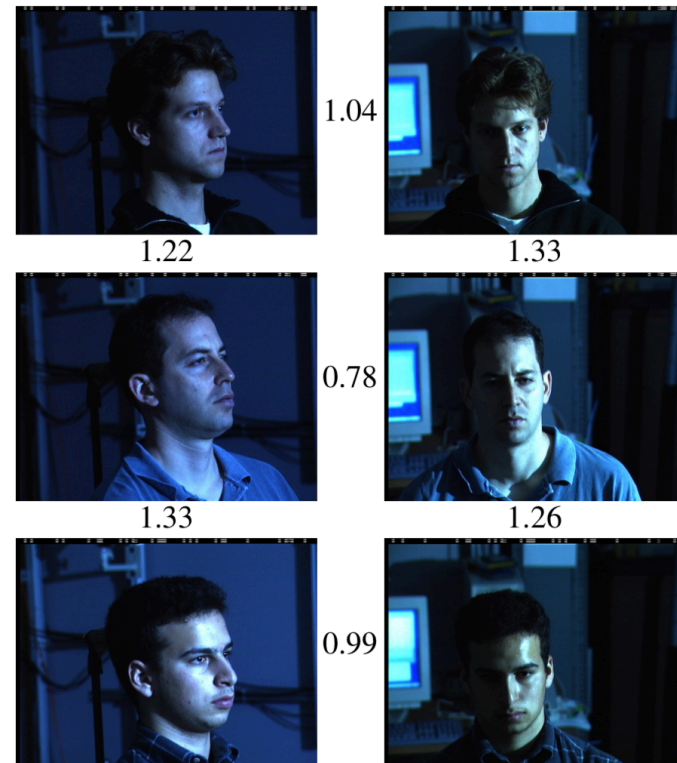


Op Meeting:

CANDLE prediction
analysis notebook

Perceptual distance vs data distance

- Metric learning
- Representation learning
- Feature encoding
- Embedding
- WordVec, ProteinVec



Siamese network



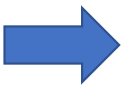
Feature Extraction | Join | Matching

CNN

CNN

+

FC Layers



Distance

Pairs could be gene expression replicates or samples from the same cancer type

Focusing on the difficult parts

- Mine the difficult samples



Figure 2. **Model structure.** Our network consists of a batch input layer and a deep CNN followed by L_2 normalization, which results in the face embedding. This is followed by the triplet loss during training.

- Change loss function

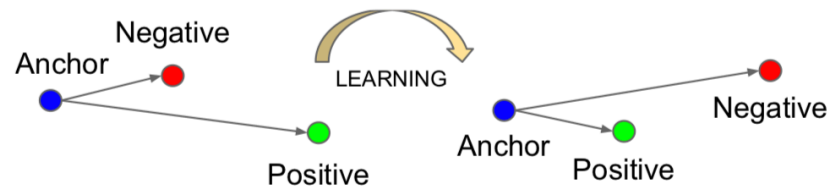
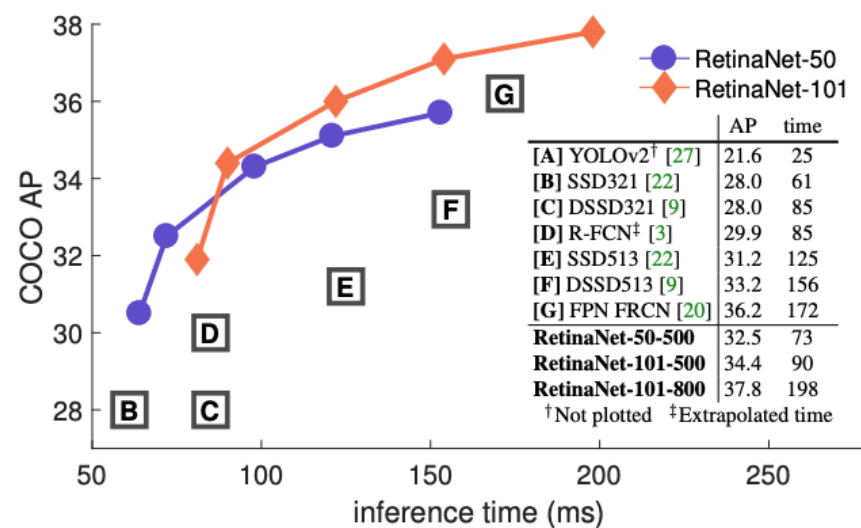
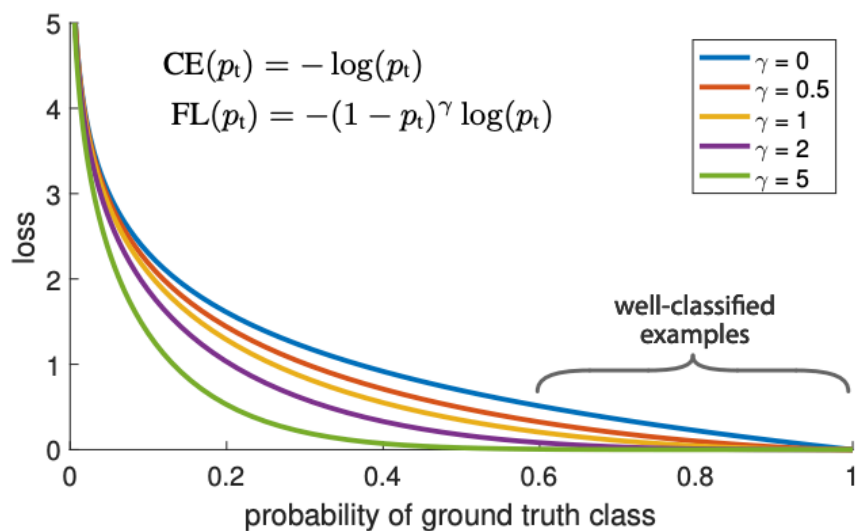


Figure 3. The **Triplet Loss** minimizes the distance between an *anchor* and a *positive*, both of which have the same identity, and maximizes the distance between the *anchor* and a *negative* of a different identity.

Focal Loss for Dense Object Detection

Tsung-Yi Lin Priya Goyal Ross Girshick Kaiming He Piotr Dollár
 Facebook AI Research (FAIR)



DL Challenge:

*from surface pattern recognition
to deep mechanistic understanding*

Just fancy regression?

Sequence to sequence models

Input	Output
1+1 =	2
19+28 =	47
577+45 =	622
👍+👍 =	✌️
19+28 =	47
五七七+四五 =	六二二

Function approximation

```
#include<stdio.h>
void main(int argc, char *argv[])
{
    int k,r;
    int i=0,j=1,f;
    int sum=1;

    r=10;
    for (k=2; k<r; k++) {
        f=i+j;
        i=j;
        j=f;
        sum=sum+j;
    }

    printf("%d\n", sum);
}
```

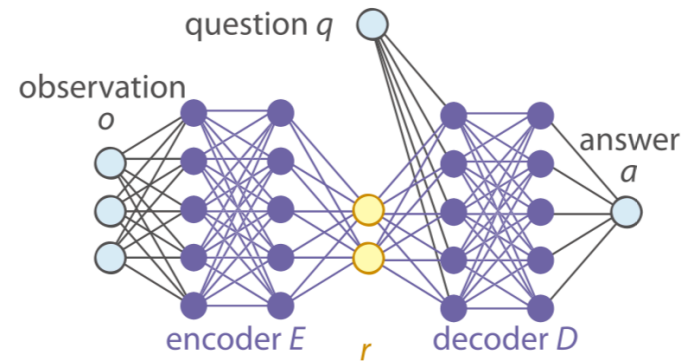
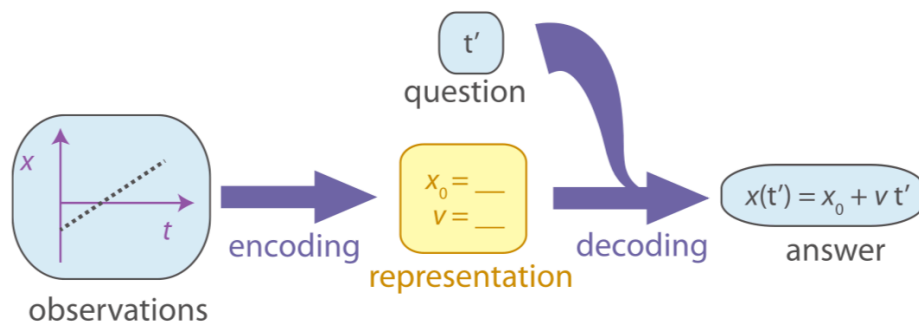
=> 88

Discovering physical concepts with neural networks

Raban Iten, Tony Metger, Henrik Wilming, Lidia del Rio, Renato Renner

(Submitted on 26 Jul 2018 (v1), last revised 29 Sep 2018 (this version, v2))

We introduce a neural network architecture that models the physical reasoning process and that can be used to extract simple physical concepts from experimental data without being provided with additional prior knowledge. We apply the neural network to a variety of simple physical examples in classical and quantum mechanics, like damped pendulums, two-particle collisions, and qubits. The network finds the physically relevant parameters, exploits conservation laws to make predictions, and can be used to gain conceptual insights. For example, given a time series of the positions of the Sun and Mars as observed from Earth, the network discovers the heliocentric model of the solar system – that is, it encodes the data into the angles of the two planets as seen from the Sun. Our work provides a first step towards answering the question whether the traditional ways by which physicists model nature naturally arise from the experimental data without any mathematical and physical pre-knowledge, or if there are alternative elegant formalisms, which may solve some of the fundamental conceptual problems in modern physics, such as the measurement problem in quantum mechanics.



JUDEA PEARL
WINNER OF THE TURING AWARD
AND DANA MACKENZIE

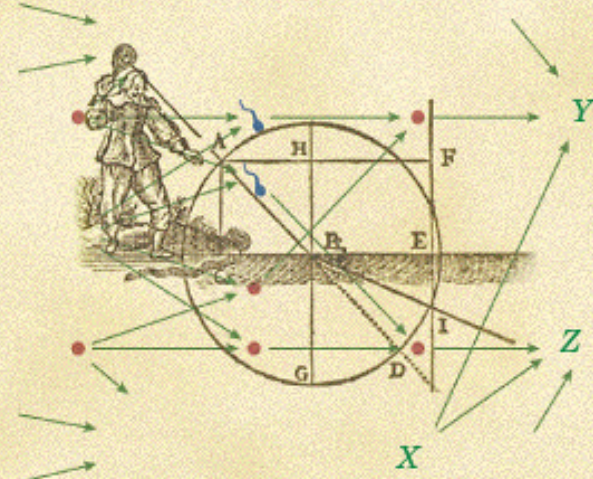
THE BOOK OF WHY



THE NEW SCIENCE
OF CAUSE AND EFFECT

CAUSALITY

MODELS, REASONING,
AND INFERENCE



JUDEA PEARL

Thank you